

**U.S. FISH AND WILDLIFE SERVICE  
SPECIES ASSESSMENT AND LISTING PRIORITY ASSIGNMENT FORM**

SCIENTIFIC NAME: *Rana luteiventris*

COMMON NAME: Columbia spotted frog (Great Basin Distinct Population Segment)

LEAD REGION: Region 8

INFORMATION CURRENT AS OF: April 15, 2009

**STATUS/ACTION**

☐ Species assessment - determined we do not have sufficient information on file to support a proposal to list the species and, therefore, it was not elevated to Candidate status

☐ New candidate

☒ Continuing candidate

☐ Non-petitioned

☒ Petitioned - Date petition received: May 1, 1989

☒ 90-day positive - FR date: October 17, 1989 (54 FR 42529)

☒ 12-month warranted but precluded - FR date: April 23, 1993 (58 FR 27260)

☐ Did the petition request a reclassification of a listed species?

**FOR PETITIONED CANDIDATE SPECIES:**

a. Is listing warranted (if yes, see summary of threats below)? YES

b. To date, has publication of a proposal to list been precluded by other higher priority listing actions? YES

c. If the answer to a. and b. is "yes", provide an explanation of why the action is precluded. Higher priority actions.

On May 1, 1989, the U.S. Fish and Wildlife Service (Service) received a petition from the Board of Directors of the Utah Nature Study Society to add the spotted frog (*Rana pretiosa*) to the List of Threatened and Endangered Species (Utah Nature Study Society 1989, pp. 1-12). The petition cited various reasons why the species should be listed including: 1) habitat destruction; 2) exotic species; 3) underfunding of both State and Federal agencies; 4) politics and conservation; and 5) large water projects such as the Central Utah Project (Utah Nature Study Society 1989, pp. 4-8). We prepared a 90-day finding on October 17, 1989, and found the petition presented substantial information that the requested action may be warranted (54 FR 42529). On May, 7, 1993, we announced our 12-month finding and found that listing the spotted frog as threatened in some portions of its range is warranted but precluded by other higher priority actions (58 FR 27260).

Another petition received in May 2004 to list all 225 candidate species, including *Rana luteiventris* as an endangered species, under the Endangered Species Act (ESA) was largely based on the present or threatened destruction, modification, or curtailment of its habitat or range, disease or predation, the inadequacy of existing regulatory mechanisms, and other natural or manmade factors affecting its continued existence (Center for Biological Diversity (CBD) *et*

al. 2004). In addition, the petitioners stated that these species have been on the candidate list for an average of 17 years and such delays have contributed to the extinction of many non-listed species (CBD *et al.* 2004). We considered the 2004 petition in this assessment; however, no new substantive information on *R. luteiventris* was presented. Two conservation agreements and strategies (Nevada Department of Wildlife (NDOW) 2003a, pp. 1-43; 2003b, pp. 1-51) were signed by Federal, State, County, and university representatives on September 30, 2003, for the Toiyabe Mountains and Northeast (Jarbidge-Independence Range and Ruby Mountains) subpopulations.

We find that the immediate issuance of a proposed rule and timely promulgation of a final rule for this species has been, since publication of the last Candidate Notice of Review (CNOR), and continues to be, precluded by higher priority listing actions (including candidate species with lower Listing Priority Numbers (LPN)) because most of our national listing budget has been consumed by work on various listing actions to comply with court orders and court-approved settlement agreements, meeting statutory deadlines for petition findings or listing determinations, emergency listing evaluations and determinations, and essential litigation-related, administrative, and program management tasks. We will continue to monitor the status of this species as new information becomes available. This review will determine if a change in status is warranted, including the need to make prompt use of emergency listing procedures. For information on listing actions taken, see the discussion of "Progress on Revising the Lists" in the current CNOR, which can be viewed on our Internet website (<http://endangered.fws.gov/>).

\_\_\_ Listing priority change

Former LPN: \_\_\_

New LPN: \_\_\_

Date when the species first became a Candidate (as currently defined): April 23, 1993

\_\_\_ Candidate removal: Former LPN: \_\_\_

\_\_\_ A – Taxon is more abundant or widespread than previously believed or not subject to the degree of threats sufficient to warrant issuance of a proposed listing or continuance of candidate status.

\_\_\_ U – Taxon not subject to the degree of threats sufficient to warrant issuance of a proposed listing or continuance of candidate status due, in part or totally, to conservation efforts that remove or reduce the threats to the species.

\_\_\_ F – Range is no longer a U.S. territory.

\_\_\_ I – Insufficient information exists on biological vulnerability and threats to support listing.

\_\_\_ M – Taxon mistakenly included in past notice of review.

\_\_\_ N – Taxon does not meet the ESA's definition of "species."

\_\_\_ X – Taxon believed to be extinct.

ANIMAL/PLANT GROUP AND FAMILY: Amphibians, Ranidae (Frogs)

HISTORICAL STATES/TERRITORIES/COUNTRIES OF OCCURRENCE: Nevada, Oregon, Idaho

**CURRENT STATES/COUNTIES/TERRITORIES/COUNTRIES OF OCCURRENCE:** Nevada (Elko, Eureka, and Nye Counties); Oregon (Lake, Harney, and Malheur Counties); Idaho (Owyhee County)

**LAND OWNERSHIP:** An estimated 90 percent of all known habitat for Columbia spotted frog (Great Basin Distinct Population Segment (DPS)) occurs on lands managed by the Forest Service and Bureau of Land Management (BLM). The Humboldt-Toiyabe National Forest (HTNF) in Nevada is the only national forest which has occupied Columbia spotted frog habitat. Occupied habitat on BLM managed lands include the Elko and Battle Mountain District Offices in Nevada; Lakeview, Burns, and Vale Field Offices in Oregon; and Jarbidge, Bruneau, and Owyhee Field Offices in Idaho. The Malheur National Wildlife Refuge in south central Oregon currently has a small population. Columbia spotted frogs are known to occur on the Yomba-Shoshone Reservation in central Nevada and the Duck Valley Reservation straddling the border of Nevada and Idaho. The State of Idaho manages a 275 hectare (680 acre) parcel at Sam Noble Springs which Columbia spotted frogs occupy. The remainder of known or suspected occupied sites occur on private lands.

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## **BIOLOGICAL INFORMATION**

### Species Description

Ranids typically are characterized as slim-waisted, long-legged, smooth-skinned jumpers with webbed hind feet and usually with a pair of dorsolateral folds (glandular folds) that extend from behind the eyes to the lower back (Figures 1-4). Adult Columbia spotted frogs measure approximately 5.6 centimeters (cm) (2.2 inches (in)) from snout to vent, with females being larger than males (Stebbins 2003, pp. 66, 229-230). Dorsal color and pattern include a light brown, dark brown, or gray, with small spots (Stebbins 2003, pp. 66, 229-230) (Figures 1-4). Ventral coloration can differ among geographic population units and may range from yellow to salmon; however, very young individuals may have very pale, almost white, ventral surfaces (Stebbins 2003, pp. 66, 229-230) (Figures 1-4). The throat and the ventral region are sometimes mottled (Figures 1-3). The head may have a dark mask with a light stripe on the upper jaw, and the eyes are turned slightly upward (Figures 1-4). Male frogs have swollen thumbs with darkened bases (Stebbins 2003, pp. 66, 229-230).

### Taxonomy

Spotted frogs (*Rana pretiosa*) were first described by Baird and Girard in 1853 (pp. 378-379) and later split into two subspecies *R. pretiosa pretiosa* and *R. pretiosa luteiventris* (Thompson 1913, pp. 53-56). The Service accepts species-specific genetic and geographic differences in



Figures 1 and 2. Columbia spotted frogs (*Rana luteiventris*) from central Nevada (Joel Sartore/joelsartore.com).



Figures 3 and 4. Columbia spotted frogs (*Rana luteiventris*) from central Nevada (Joel Sartore/[joelsartore.com](http://joelsartore.com)).

Columbia spotted frogs based on previous work by Green *et al.* (1996, pp. 377-388; 1997, pp. 2-7), Bos and Sites (2001, pp. 1505-1511), and more recently by Funk *et al.* (2008, pp. 201-202) which define populations in western Washington and Oregon and northeastern California as Oregon spotted frogs (*R. pretiosa*) and the remainder of the populations as Columbia spotted frogs (*R. luteiventris*) (Figure 5). Based on further geographic and genetic characterization, Columbia spotted frogs in southwest Idaho, southeast Oregon, and northeast and central Nevada are part of the Great Basin population of Columbia spotted frogs (Funk *et al.* 2008, pp. 201-202). It was previously thought that populations in northeast Oregon were part of the Great Basin population; however, Funk *et al.* (2008, pp. 201-202) found that these populations belong to the Northern population (see Distinct Population Segment discussion below). A small population on the eastern border of White Pine County, Nevada, and Toole County, Utah, has been determined through phylogenetic data to be part of the Utah population of Columbia spotted frogs (Funk *et al.* 2008, pp. 201-202). We have carefully reviewed available taxonomic information to reach the conclusion that the species *R. luteiventris* is a valid taxon.

#### Habitat

Columbia spotted frogs are found closely associated with clear, slow-moving or ponded surface waters, with little shade, and relatively constant water temperatures (Reaser 1997, pp. 32-33; Reaser and Pilliod 2005, p. 561; Welch and MacMahon 2005, p. 477). Reproducing populations have been found in habitats characterized by springs, floating vegetation, and larger bodies of pooled water (e.g., oxbows, lakes, stock ponds, beaver-created ponds, seeps in wet meadows, backwaters) (Reaser and Pilliod 2005, p. 560). A deep silt or muck substrate may be required for hibernation and torpor (Bull 2005, p. 12; Reaser and Pilliod 2005, p. 561). In colder portions of their range, Columbia spotted frogs will use areas where water does not freeze, such as spring heads and undercut streambanks with overhanging vegetation (Bull 2005, p. 12; Reaser and Pilliod 2005, p. 561); however, they can overwinter underneath ice-covered ponds (Tattersall and Ultsch 2008, pp. 122-123). Females usually lay egg masses in the warmest areas of a pond, typically in shallow water (10-20 cm, 4-8 in), and clutch sizes vary (150-2,400 eggs) (Bull 2005, pp. 8 and 11; Reaser and Pilliod 2005, p. 560; Pearl *et al.* 2007a, pp. 87-89). Successful egg production and the viability and metamorphosis of Columbia spotted frogs are susceptible to habitat variables such as temperature, depth, and pH of water, cover, and the presence or absence of predators (Munger *et al.* 1996, p. 8; Reaser 1996, pp. 21-22; Bull 2005, p. 7; Reaser and Pilliod 2005, pp. 561-562).

#### Current and Historical Range/Distribution-Nevada

Columbia spotted frogs in Nevada are found in the central (Nye County) and northeastern (Elko and Eureka Counties) parts of the State, usually at elevations between 1,700 and 2,650 meters (5,600 and 8,700 feet), although they have been recorded historically in a broader range including Lander County in central Nevada and Humboldt County in northwest Nevada (Reaser 2000, p. 1159). The Great Basin population of Columbia spotted frogs in Nevada is geographically separated into three distinct subpopulations: Jarbidge-Independence Range, Ruby Mountains, and Toiyabe Mountains. The Nevada Wildlife Action Plan defines these areas as the Great Basin and Columbia Plateau Ecoregions (NDOW 2006, p. 66).

The largest of Nevada's three subpopulation areas is the Jarbidge-Independence Range in Elko



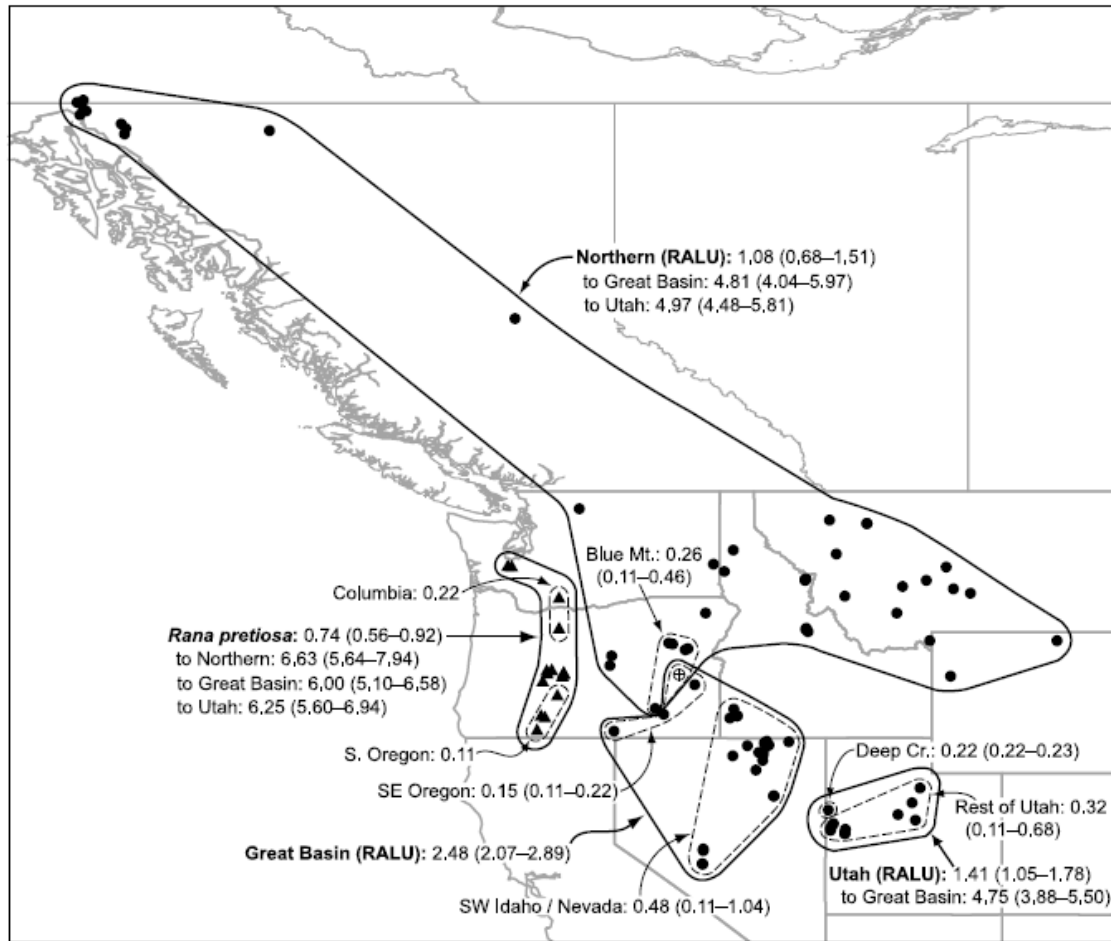


Fig. 3. Geographic distribution of major clades (solid black lines) and nested clades (dashed lines) identified in the phylogenetic analyses, with mean percent corrected sequence divergence (and ranges in parentheses) shown within and among clades. Clade names correspond to those used in Fig. 2 and Tables 1–3. Triangles = *R. pretiosa*; circles = *R. luteiventris*. The open circle with a cross is Kingsbury Gulch (site 59), where haplotypes from the northern clade and Great Basin clade were found.

Figure 5. Geographic distribution of Oregon spotted frogs (*Rana pretiosa*) and Columbia spotted frogs (*Rana luteiventris*) (Funk *et al.* 2008, p. 202). Reprinted with permission.

and Eureka Counties. This subpopulation area is formed by the headwaters of streams in two major hydrographic basins. The South Fork Owyhee River, Owyhee River, Bruneau River, and Salmon Falls Creek drainages flow north into the Snake River basin. Mary's River, North Fork Humboldt River, and Maggie Creek drain into the interior Humboldt River basin. Columbia spotted frogs occur in the Ruby Mountains in tributaries to the South Fork Humboldt River including Green Mountain, Smith, Corral, and Rattlesnake Creeks on lands in Elko County managed by the HTNF. In the Toiyabe Mountains, Columbia spotted frogs are found in seven drainages in Nye County, Nevada--the Reese River (Upper and Lower), Cow and Ledbetter Canyons, and Cloverdale, Stewart, Illinois, and Indian Valley Creeks (NDOW 2003b, p. S-8). The Toiyabe Mountains subpopulation is geographically isolated from the Ruby Mountains and Jarbidge-Independence Range subpopulations by a large gap in suitable habitat and represents the southern-most extremity of the species' range.

### Current and Historical Range/Distribution-Idaho and Oregon (Owyhee subpopulation)

Prior to 1995, only six historical sites were known in the Owyhee Mountain range in Idaho (Munger *et al.* 1996, pp. 2-3, 16) and only 22 sites were known in southeastern Oregon in Malheur County (Munger *et al.* 1998, pp. 6-7). Currently, Columbia spotted frogs appear to be widely distributed throughout southwestern Idaho (Owyhee County) and southeastern Oregon, but local populations within this general area appear to be isolated from each other by either natural or human-induced habitat disruptions (Smyth 2004, pp. 3-7; Bull 2005, pp. 2-3; Engle 2006, p. 20; Moser and Patton 2006, p. 7). The Idaho Comprehensive Wildlife Strategy defines this area as the Owyhee Uplands (Idaho Department of Fish and Game (IDFG) 2005, pp. 1-8). In southeastern Oregon, the historical and current range of Columbia spotted frogs include, but are not limited to, the Owyhee and Steens Mountains in Harney and Malheur Counties (Munger *et al.* 1998, pp. 3-4; Smyth 2004, pp. 3-7; Funk *et al.* 2008, p. 202). The Oregon Conservation Strategy defines this area as the Northern Basin and Range Ecoregion (Oregon Department of Fish and Wildlife (ODFW) 2006, pp. 204-221).

### Population Estimates/Status

Status-Nevada: Declines of Columbia spotted frog populations in Nevada have been recorded since 1962 when it was observed that in many Elko County localities where Columbia spotted frogs were once numerous, the species was nearly extirpated (Turner 1962, pp. 326-327). Extensive loss of habitat was found to have occurred from conversion of wetland habitats to irrigated pasture and from spring and stream dewatering by mining and irrigation practices. In addition, there was evidence of extensive impacts on riparian habitats due to intensive livestock grazing. Researchers in Nevada have documented the loss of historically occupied sites, reduced numbers of individuals within local populations, and declines in the reproduction of those individuals (Turner 1962, pp. 326-327; Hovingh 1990, p. 6; Reaser 1997, pp. 30-33; Hatch *et al.* 2002, pp. 47-50; Wentz *et al.* 2005, p. 99). Between 1994 and 1996, Reaser (1997, pp. 30-31) resurveyed 41 (45 percent) of 91 previously occupied sites identified between 1912 and 1992. Of the 41 previously occupied sites visited, 14 (34 percent) were still occupied while 27 (66 percent) were unoccupied (Reaser 1997, pp. 30-31).

Between 2002 and 2006, Forest Service crews in northeastern Nevada resurveyed previously surveyed sites that were identified during the 1993-1998 efforts by Reaser and others (Amy 2003, pp. 1-6). Of the 625 sites visited, Columbia spotted frogs were present at 136 sites (22 percent) and were not detected at the remaining 489 sites (78 percent) (Amy 2003, p. 2; 2004, p. 2; Meneks 2005a, p. 3; 2005b, p. 5; 2006, p. 7). Within the Ruby Mountains, Jarbidge, and Mountain City Ranger Districts and the BLM Elko District Office in northeast Nevada, there are approximately 235 6<sup>th</sup> code hydrologic units (HUCs). From 2000 to present, the Forest Service and NDOW have conducted presence-absence surveys in 75 HUCs (32 percent) and have detected Columbia spotted frogs in 44 percent (33 HUCs) of the HUCs sampled (J. Petersen, NDOW, pers. comm. 2009). Additionally, presence-absence surveys were conducted by the Service and Tribal members on the Nevada portion of the Duck Valley Tribal lands during 2004 and 2005, where the species was found in seven out of 16 locations surveyed (Service, unpublished data).



In 2004, the Forest Service initiated an intensive mark-recapture survey at two sites, Green Mountain Creek, Ruby Mountains Ranger District and Tennessee Gulch, Mountain City Ranger District (and added a third site in 2005, Pole Creek, Jarbidge Ranger District), as part of an effort to determine population estimates, mortality, juvenile-to-adult recruitment, movement, and habitat preference (Meneks 2005a, pp. 1-3). Between 2004 and 2008, a total of 1,889 frogs were captured, 1,527 of which were marked using Passive Integrated Transponder (PIT) tags from all three sites (Meneks 2008, pp. 2-11). Between 2006 and 2008, the number of adult frogs captured at the Green Mountain Creek site seem stable and remain double the numbers captured from 2004 and 2005; however, juvenile numbers have shown a more variable trend (Meneks 2008, pp. 3-4). Adult numbers captured at Tennessee Gulch between 2005 and 2008 were similar and were three times greater than the number captured during the 2004 survey, while juvenile numbers remained low for the second year in a row (Meneks 2008, pp. 4-6). The number of adult frogs captured at the Pole Creek site between 2005 and 2007 averaged 200 individuals; however, in 2008, 114 adults were captured. Additionally, juvenile numbers remained low with a total of eight juveniles captured between 2006 and 2008 (0 in 2008) compared to 72 captured in 2005 (Meneks 2008, pp. 6-9).

During the summers of 2000 and 2001, mark-recapture surveys of the Toiyabe Mountains subpopulation were conducted by the University of Nevada, Reno. Preliminary estimates of frog numbers in the Indian Valley Creek drainage were approximately 5,000 breeding individuals, which was greater than previously believed (Hatch, *et al.* 2002, p. 3). However, during the 2000-2001 winter, Hatch *et al.* (2002, p. 23) noted a large population decrease, ranging between 66 and 86.5 percent at several sites. Survey results suggested poor winter habitat contributed to the winterkill (Hatch *et al.* 2002, pp. 25-27). A large mark-recapture study using PIT tags was initiated for the Toiyabe Mountains subpopulation in 2004 and has continued annually. During this period, approximately 1,850 frogs have been PIT tagged. Results from the 2008 monitoring are discussed below (NDOW 2009a, pp. 4-6). Total adult frog captures were similar in 2008 ( $n = 628$ ) and 2007 ( $n = 674$ ) and are substantially higher than in previous years. Total recaptures in 2008 were the highest recorded since monitoring began with 226 individuals captured from previous years (NDOW 2009a). Juvenile frog counts in 2008 ( $n = 634$ ) and 2007 ( $n = 646$ ) were similar; however, numbers captured were substantially higher than the 2004 ( $n = 68$ ), 2005 ( $n = 92$ ), and 2006 ( $n = 251$ ) surveys. Population estimates derived from the Jolly-Seber method calculated an adult population of 2,189 (95 percent confidence interval 1,860-2,827) frogs for the seven sentinel site locations combined in 2007, compared to 2,029 (95 percent confidence interval 1,693-2,683) adults in 2006 and 1,421 (95 percent confidence interval 1,190-1,870) adults in 2005.

The lack of standardized and extensive monitoring and routine surveying has prevented dependable determinations of frog population numbers or trends across Nevada. However, since the signing of a conservation agreement and strategy (CAS) in 2003 (NDOW 2003a, b), standardized protocols and consistent monitoring are taking place in both the Northeast (Jarbidge-Independence Range and Ruby Mountains) and Toiyabe Mountains subpopulations. A long-term monitoring plan to standardize monitoring locations and protocols has also been developed for the Toiyabe Mountains subpopulation (NDOW 2004b, pp. 1-25) and has been

implemented annually since 2004.

Status-Idaho: Extensive surveys since 1996 throughout southwestern Idaho have increased the number of known spotted frog sites. However, most of these surveys suggest the sites support small numbers of frogs. Additionally, all known local populations in southwestern Idaho appear to be functionally isolated (Engle 2001, p. 3; Engle and Munger 2003, pp. 3-11). Surveys conducted in 2001 reported that of the 49 known local populations in southwestern Idaho, 61 percent had five or fewer adult frogs (Engle 2002, p. 3). The largest known local population of Columbia spotted frogs occurs at Sam Noble Springs in the Rock Creek drainage of Owyhee County. Monitoring of the adult frog population has occurred annually since 1998 and no more than 150 adult frogs have been captured in any one year (Lohr and Moser 2008, p. 10). Despite the inability to estimate population size in some years due to inconsistencies in data collection, it appears that the adult spotted frog population at Sam Noble Springs suffered a brief decline in 2003, but data collected from 2005 to 2008 show an adult population between 80 and 100 individuals (Lohr and Moser 2008, p. 11). Additionally, in 2008, the number of egg masses observed was the highest since monitoring began in 2000 (Lohr and Moser 2008, p. 13).

Extensive monitoring at sentinel sites between 2000 and 2002 indicated a 36 percent decline in the number of adult Columbia spotted frogs encountered (Lingo and Munger 2003, p. 26). The overall population at one sentinel site, Stoneman Creek, has increased partially due to habitat improvements (Munger and Oelrich 2006, p. 8; Lohr and Moser 2008, p. 13). Continued annual monitoring at sentinel sites is needed to understand population fluctuations and to document trends. Boise State University has conducted several research projects related to spotted frogs including the reintroduction of beaver (*Castor canadensis*) for Columbia spotted frog habitat restoration (Munger and Lingo 2003, pp. 1-6), effects of grazing (Howard and Munger 2003, pp. 9-13), spotted frog habitat evaluations (Munger 2003, pp. 4-12), and sentinel site surveys (Lingo and Munger 2003, pp. 1-69; Blankinship and Munger 2005, pp. 1-65; Munger and Oelrich 2006, pp. 1-19).

In 2007 and 2008 a proportion of area occupied study was implemented in Owyhee County as a method to obtain a better understanding of the species' trend as it relates to occupancy of catchment basins (Moser 2007, pp. 9-10; Lohr and Moser 2008, p. 9). Of the 31 catchment basins surveyed in 2008, 21 (68 percent) had historical occurrence of spotted frogs (Lohr and Moser 2008, p. 14). Of those with historical occurrences, 18 (86 percent) were occupied by spotted frogs in 2008 (Lohr and Moser 2008, p. 14). Spotted frogs occupied about 62 percent of the pilot study area in 2008 (Lohr and Moser 2008, p. 14).

Status-Oregon: In southeastern Oregon, surveys conducted in 1997 reconfirmed a population of Columbia spotted frogs in the Dry Creek drainage in Malheur County (Munger *et al.* 1998, pp. 3-4). Detailed population estimates using PIT tags have occurred in Dry Creek since 2001 (Meyer 2008, pp. 1-125). Results from these efforts suggest a fairly large reproducing population exists in this area and all life stages have generally increased since 2001; however, survival rates are low (Meyer 2008, p. 33). The United States Geological Survey (USGS) has performed annual monitoring of the Kingsbury Gulch site since 2002, and they have documented a sharp decline in this population most likely due to habitat alteration (Adams *et al.* 2008, pp. 10-11). Presence-

absence monitoring has occurred in the Steens Mountains area, Harney County, in which small isolated populations of Columbia spotted frogs have been located (Smyth 2004, pp. 3-7).

Between 2000 and 2003, the USGS compared current regional distributions of amphibians with occurrence patterns suggested in historical data (Adams *et al.* 2006, pp. 1-21). Visual encounter surveys were used to determine presence-absence of Columbia spotted frogs on public lands in eastern Oregon and northern Nevada. Based on occupancy models, USGS estimated that Columbia spotted frogs occupied 53 percent of the 30 historical sites in the area surveyed (Wente *et al.* 2005, p. 99). Six of 16 sites proximal to historical sites were occupied between 2000 and 2003 (Wente *et al.* 2005, p. 99). Additionally, 187 sites in southeastern Oregon were randomly selected for presence-absence surveys of which only three sites were occupied; however, variability in occupancy between the 3 years was problematic (Wente *et al.* 2005, pp. 99-106).

In summary, monitoring efforts are being implemented throughout the range of the Columbia spotted frog in Idaho, Nevada, and Oregon; however, lack of consistency in survey protocols and monitoring efforts make it difficult to understand the status of the species across its range. Furthermore, deciphering historical data collected throughout the 1900's and comparing it to current occupancy rates has been problematic. A range-wide effort to determine historical and current occupancy is needed to better track the status of this species.

#### **DISTINCT POPULATION SEGMENT (DPS)**

Under the ESA, we must consider for listing any species, subspecies, or, for vertebrates, DPSs of these taxa, if information is sufficient to indicate that such action may be warranted. To implement the measures prescribed by the ESA and its Congressional guidance, we, along with the National Oceanic and Atmospheric Administration (NOAA) Fisheries, developed policy to clarify our interpretation of the phrase “distinct population segment of any species of vertebrate fish or wildlife” for the purposes of listing, delisting, and reclassifying species under the ESA (February 7, 1996, 61 FR 4722). The policy allowed us to interpret the requirement of the ESA to “...determine whether any species is an endangered species or a threatened species” (section 4(a) (1)) in a clear and consistent fashion for the term “distinct population segment.” Under our DPS policy, we consider three elements in a decision regarding the status of a possible DPS as endangered or threatened under the ESA. These are applied similarly for addition to the lists of endangered and threatened wildlife and plants, for reclassification, and for removal. The elements are: (1) the population segment's discreteness from the remainder of the species to which it belongs; (2) the population segment's significance to the species to which it belongs; and (3) the population segment's conservation status in relation to ESA standards for listing (i.e., when treated as if it were a species, is the population segment endangered or threatened?). Our policy further recognizes it may be appropriate to assign different classifications to different DPSs of the same vertebrate taxon (61 FR 4722).

#### **Discreteness**

The DPS policy standard for discreteness allows an entity given DPS status under the ESA to be adequately defined and described in some way that distinguishes it from other representatives of

its species. A population segment of a vertebrate species may be considered discrete if it satisfies either one of the following two conditions: (1) it is markedly separated from other populations of the same taxon as a consequence of physical, physiological, ecological, or behavioral factors (quantitative measures of genetic or morphological discontinuity may provide evidence of this separation); or (2) it is delimited by international governmental boundaries within which significant differences in control of exploitation, management of habitat, conservation status, or regulatory mechanisms exist.

Columbia spotted frogs in Nevada, southwestern Idaho, and most populations in the southeastern Oregon portion of the Great Basin are geographically separate from the remainder of the species; however, one isolated site in southeastern Oregon (Kingsbury Gulch) showed genetic evidence of an overlap between the Northern and Great Basin populations (Funk *et al.* 2008, p. 204) (Figure 1). For management purposes, populations within the Great Basin have been divided into four subpopulations. The largest of Nevada's three subpopulation areas is the Jarbidge-Independence Range in Elko and Eureka Counties. This subpopulation area is formed by the headwaters of streams in two major hydrographic basins. The South Fork Owyhee River, Owyhee River, Bruneau River, and Salmon Falls Creek drainages flow north into the Snake River basin. Marys River, North Fork of the Humboldt River, and Maggie Creek drain into the interior Humboldt River basin. A smaller subpopulation of Columbia spotted frogs is located in the Ruby Mountains about 80 kilometers (km) (50 miles (mi)) south of the Jarbidge-Independence Range subpopulation. However, these two subpopulations are isolated by lack of suitable habitat and hydrologic connectivity. The Toiyabe Mountains subpopulation is isolated nearly 320 km (200 mi) southeast of the Ruby Mountains and Jarbidge-Independence Range subpopulations and represents the southern-most extremity of its range. The Owyhee subpopulation of Columbia spotted frogs appears to be widely distributed throughout southwestern Idaho (Owyhee County) and southeastern Oregon (Lake, Malheur, and Harney Counties), but local populations within this general area are small and appear to be isolated from each other and from subpopulations in northeastern Nevada by either natural or human-induced habitat disruptions.

All of these Great Basin subpopulations are geographically isolated and separate from the main continuous population of Columbia spotted frogs in the central mountains of Idaho by the Snake River Plain and adjacent lowlands in eastern Oregon. The Owyhee subpopulation in southwestern Idaho is approximately 160 km (100 mi) from the main continuous population in central Idaho. Occupied habitat in the main population is characterized by conifer forests and high elevation lake environments while habitat for the Great Basin population is characterized by sagebrush with stream and pond environments. Furthermore, the Great Basin population is both hydrologically and geographically separated from isolated populations in Utah. The subpopulation in the Ruby Mountains (Lahontan Basin) is approximately 145 km (90 mi) from the West Desert population (Bonneville Basin) near Ibapah, Utah. As detailed below, geographic isolation of the Great Basin population is supported by genetic analyses.

Three earlier genetic studies were conducted on Columbia spotted frogs which have improved our knowledge on the distribution and genetic structure of the species (Green *et al.* 1996, pp. 374-390; Green *et al.* 1997, pp. 1-8; Bos and Sites 2001, pp. 1499-1513). Unfortunately, these

studies did not adequately sample populations in southwestern Idaho and eastern Oregon. Because the distribution of distinct subpopulations within the Great Basin DPS was unresolved, the USGS initiated a genetic evaluation of the Great Basin DPS (USGS 2006, pp. 1-3). Objectives of the study included: 1) determine the distribution of distinct subpopulations within the Great Basin DPS; 2) determine whether Columbia spotted frog populations from southeastern Oregon and southern Idaho are part of the Great Basin DPS; 3) determine whether Columbia spotted frog populations from northeastern Oregon are part of the Great Basin DPS or instead, part of the large, contiguous portion of the species' range in the northern Rocky Mountains; and 4) examine population genetic structure and status in the Great Basin DPS of Columbia spotted frog. Results from this study are presented below.

The strongest genetic evidence that Great Basin frogs are genetically discrete from other Columbia spotted frogs comes from Funk *et al.* (2008, pp. 198-210) who examined mitochondrial DNA (mtDNA) sequence variation throughout the extant range of Columbia spotted frogs. These data indicate three distinct major clades (a clade is a group of taxa sharing a closer common ancestry with one another than with members of any other clade); Northern, Great Basin, and Utah (Funk *et al.* 2008, pp. 201-202) (Figure 5). The three clades are nearly as divergent from each other as they are from Oregon spotted frog, a closely related but separate species (Funk *et al.* 2008, p. 202). Additionally, within each major clade, well-defined nested clades are also evident. The Great Basin clade has two well-defined nested clades in southwestern Idaho-Nevada and southeastern Oregon (Funk *et al.* 2008, p. 202) (Figure 5). These two nested clades are also the most divergent among the nested clades indicating the effects of small isolated populations in southeastern Oregon (Funk *et al.* 2008, p. 205). The authors also found one location in southeastern Oregon in which there is an overlap between the Northern and Great Basin clades (Funk *et al.* 2008, p. 204) (Figure 5). This area of southeastern Oregon has been identified as a natural zone of hybridization for other species (Remington 1968, pp. 321-428). More genetic analyses will be conducted in southeastern Oregon in 2009 to further define the phylogeographic break between the Northern and Great Basin populations.

## **Significance**

Under our DPS policy, once we have determined that a population segment is discrete, we consider its biological and ecological significance to the larger taxon to which it belongs. This consideration may include, but is not limited to, evidence of the persistence of the discrete population segment in an ecological setting that is unique for the taxon; evidence that loss of the population segment would result in a significant gap in the range of the taxon; evidence that the population segment represents the only surviving natural occurrence of a taxon that may be more abundant elsewhere as an introduced population outside its historical range; and evidence that the discrete population segment differs markedly from other populations of the species in its genetic characteristics.

We have found substantial evidence that two of these significance factors are met by the Great Basin population of the Columbia spotted frog. The extinction of the Nevada, southwestern Idaho and southeastern Oregon portion of the range of the Columbia spotted frog would likely result in the loss of a significant genetic entity and the curtailment of the range of the species.

Particularly, the work of Funk *et al.* (2008, pp. 198-210) indicates that Columbia spotted frogs in the Great Basin differ genetically from Columbia spotted frogs sampled in other portions of the range to a significant degree. Additionally, loss of Columbia spotted frogs in Nevada, southwestern Idaho and southeastern Oregon would eliminate the southern extent of the species' range.

## **Conclusion**

We evaluated the Great Basin population of Columbia spotted frogs, addressing the two elements which our policy requires us to consider in deciding whether a vertebrate population may be recognized as a DPS and considered for listing under the ESA. We conclude that the Great Basin population is discrete, as per our policy, based on its geographic separation and genetic divergence from the isolated populations in Utah and the main continuous populations in central and northern Idaho, northeastern Oregon, eastern Washington, western Montana, northwestern Wyoming, and southeast Alaska, and British Columbia and Alberta, Canada. We conclude that the Great Basin population of the Columbia spotted frog is significant because the loss of the species from this area would result in a significant reduction in the species' range and would constitute loss of a genetically divergent portion of the species. Because the population segment meets the discreteness and significance criteria of our DPS policy, the Great Basin population of the Columbia spotted frog constitutes a DPS which qualifies for consideration for listing.

## **THREATS**

### **A. The present or threatened destruction, modification, or curtailment of its habitat or range.**

Habitat modification and/or destruction has been implicated in the majority of amphibian declines (Bishop *et al.* 2003, pp. 209-210; Young *et al.* 2004, pp. 31-32; Bradford 2005, pp. 919, 921-922; Vredenburg and Wake 2007, p. 5; Wells 2007, pp. 817-825; Stuart *et al.* 2008, pp. 39-42). Isolated populations of amphibians, as seen throughout the range of Columbia spotted frogs in the Great Basin, are particularly susceptible to habitat modification (Noss *et al.* 2006, p. 230; Tait 2007, p. 26). Columbia spotted frog habitat degradation and fragmentation is a combined result of past and current land use influences from agricultural development, intensive livestock grazing, spring development, urbanization, mining activities, and climate change. Small upland streams and meadows found throughout the central Great Basin are inherently unstable and have been prone to incision for at least the last 400-500 years (Germanoski and Miller 2004, p. 117). Land use activities in these sensitive areas have initiated or accelerated the incision process which has changed the hydrologic function of meadow systems (Jewett *et al.* 2004, pp. 152-155). These changes in the hydrology of meadows, mainly the lowering of the water table, can cause the vegetation communities to shift from wet meadow communities (*Carex* sp.) to dry upland plant communities (*Artemisia* sp.) (Chambers *et al.* 2004a, pp. 201-205). The loss of meadow complexes limits the available habitat for Columbia spotted frogs to the incised channel which may cause a crowding effect (Noss *et al.* 2006, p. 223). Natural fluctuations in environmental conditions (i.e., drought) tend to magnify the detrimental effects of land use activities, just as the land use activities may compound the detrimental effects of natural environmental events (Boone *et al.* 2003, pp. 138-142).

Fragmentation of habitat may be one of the most significant barriers to Columbia spotted frog recovery and population persistence (Semlitsch 2002, pp. 620-623; Green 2003, pp. 340-341; Opdam and Wascher 2004, pp. 285-297; Funk *et al.* 2005a, pp. 14-15; Tait 2007, p. 26). Recent studies in Idaho indicate that Columbia spotted frogs exhibit breeding site fidelity (Pilliod *et al.* 2002, pp. 1853-1859; Engle and Munger 2003, pp. 9-10). Movement of frogs from hibernation ponds to breeding ponds may be impeded by zones of unsuitable habitat which can lead to local population extinctions (Engle and Munger 2003, pp. 12-13; Funk *et al.* 2005a, p. 15; Funk *et al.* 2005b, p. 494). As movement corridors become more fragmented through loss of flows within riparian or meadow habitats, local populations will become more isolated (Bull and Hayes 2001, pp. 120-122; Pilliod *et al.* 2002, pp. 1853-1859; Engle and Munger 2003, pp. 12-13; Munger 2003, pp. 4-9; Funk *et al.* 2005a, p. 15; Funk *et al.* 2005b, p. 494; Semlitsch 2008, pp. 260-265). Vegetation and surface water along movement corridors provide relief from high temperatures and arid environmental conditions, as well as protection from predators. Loss of vegetation and lowering of the water table as a result of the above mentioned activities can pose a significant threat to frogs moving from one area to another. Likewise, fragmentation and loss of habitat can prevent frogs from colonizing suitable sites elsewhere (Gibbs 2000, pp. 316-317; Semlitsch 2002, pp. 621-623; Funk *et al.* 2005b, p. 494; Pringle 2006, pp. 243-246).

Springs provide a stable, permanent source of water for frog breeding, feeding, and winter refugia (IDFG *et al.* 1995, p. 9). Springs provide deep, protected areas which serve as hibernacula for Columbia spotted frogs in cold climates. Springs also provide protection from predation through underground openings (IDFG *et al.* 1995, p. 9; Patla and Peterson 1996, pp. 16-17). Analyzing 10 different threats that influence the abundance and distribution of taxa associated with spring systems in the Great Basin, Sada and Vinyard (2002, p. 280) found that spring developments were associated with the greatest number of taxa being affected. Most spring developments result in the installation of a pipe or box to fully capture the water source and direct water to another location such as a livestock watering trough. Loss of this permanent source of water in semi-arid ecosystems can also lead to the loss of associated riparian habitats and wetlands used by Columbia spotted frogs. Developed spring pools could be functioning as attractive nuisances for frogs, concentrating them into isolated groups, increasing the risk of disease and predation (Noss *et al.* 2006, p. 223). Many of the springs in southern Idaho, eastern Oregon, and Nevada have been developed for agricultural use.

According to Minshall *et al.* (1989, p. 118), riparian and stream ecosystems are the most threatened ecosystems in the Great Basin. Behnke and Zarn (1976, p. 5) identified livestock grazing as the greatest threat to the integrity of stream habitat in the western United States. Grazing occurs throughout the range of Columbia spotted frogs and has been cited as detrimental to Columbia spotted frog habitat (Munger *et al.* 1996, p. 9; Reaser 1997, pp. 37-38; Engle 2002, pp. 44-55; Service 2006, pp. 4-5). Though direct correlation between Columbia spotted frog declines and livestock grazing is limited, the effects of heavy grazing on riparian areas are well documented (Kauffman *et al.* 1983a, pp. 684-685; 1983b, pp. 686-689; Kauffman and Kreuger 1984, pp. 432-434; Schulz and Leininger 1990, pp. 297-299; Belsky *et al.* 1999, pp. 425-428).

Bull and Hayes (2000, pp. 292-294) found no impacts of cattle grazing on the reproductive



success of Columbia spotted frogs in ponds in northeastern Oregon; however, there was high variability in their results and grazing intensity and timing was not evaluated. Adams *et al.* (2009, pp. 135-137) found no significant short-term effects of cattle exclosures on the number of Columbia spotted frog egg masses, larval survival, size of metamorphs, or water quality measurements. Moreover, nutrient levels often associated with negative impacts to amphibians were very low to non-detectable (Adams *et al.* 2009, pp. 136-137). In contrast, Howard and Munger (2003, p. 10) found lower survival of Columbia spotted frog larvae in their high livestock waste treatment; however, the high waste treatment larvae had higher growth rates. Schmutzer *et al.* (2008, pp. 2617-2619) found significantly larger green frog (*R. clamitans*), bullfrog (*R. catesbeiana*), and pickerel frog (*R. palustris*) larvae in ponds with cattle grazing; however, larval abundance was significantly higher in ponds with no cattle grazing. Additionally, water quality measurements including turbidity, specific conductivity, and dissolved oxygen, were significantly higher in ponds with grazing (Schmutzer *et al.* 2008, pp. 2618-2619). In a behavioral study, Shovlain *et al.* (2005, pp. 10-12) found that Oregon spotted frogs increased their use of grazing exclosures under heavy grazing pressure while no preferences were found under a light grazing regime. Jansen and Healey (2003, pp. 211-218) found that amphibian species diversity declined and habitat condition decreased with increasing grazing intensity along a river in southeastern Australia.

The reduction of beaver populations has been noted as an important feature in the reduction of suitable habitat for Columbia spotted frogs (Reaser 1997, p. 39; ODFW 2006, p. 288). Beaver are important in the creation of small pools with slow-moving water that function as habitat for frog reproduction and create wet meadows that provide foraging habitat and protective vegetation cover (Cunningham *et al.* 2007, pp. 2520-2523; Stevens *et al.* 2007, pp. 6-11). Beaver trapping is still common in Idaho and harvest is unregulated in most areas (IDFG *et al.* 1995, p. 10). In some areas, beavers are removed because of a perceived threat to water for agriculture. As indicated above, permanent ponded waters are important in maintaining spotted frog habitats during severe drought and winter periods. Removal of beaver in 1992 and the subsequent deterioration of the beaver dam on Stoneman Creek in Idaho is believed to be directly related to the decline of a spotted frog population there (Lingo and Munger 2003, pp. 3-6; Munger and Oelrich 2006, pp. 5-8). Intensive surveying of Stoneman Creek documented only one adult Columbia spotted frog in 2000 (Engle 2000, p. 4); however, in 2001, a beaver reintroduction project was started on Stoneman Creek and by 2002 it had one of the highest frog recruitment classes in the Owyhee subpopulation (Lingo and Munger 2003, p. 5).

The effects of mining on Great Basin Columbia spotted frogs have not been specifically studied, but the adverse effects of mining activities on water quality and quantity, other wildlife species, and amphibians in particular have been addressed in professional scientific forums (Ripley *et al.* 1996, pp. 49-111; Lefcort *et al.* 1998, pp. 449-452; Burkhart *et al.* 2003, pp. 111-128; Unrine *et al.* 2004, pp. 2966-2969; Bridges and Semlitsch 2005, pp. 89-92). Mining can contribute toxic substances into waterways, alter stream morphology, and dewater streams completely (Nelson *et al.* 1991, pp. 429-446; Service 2008, pp. 30-33). Up until 2001, Nevada had the second-highest level of atmospheric mercury releases in the nation (Miller 2004, p. 1). According to Toxic Release Inventory data from the Environmental Protection Agency (USEPA), major precious metal mining facilities in Nevada released between 5,443.1 and 5,896.7 kilograms (12,000 and

13,000 pounds) of mercury directly into the atmosphere from 1998 to 2001 (Higgins *et al.* 2007, p. 3), the majority of which came from the gold mining industry (USEPA 2006, pp. 1-4). Additionally, a recent advisory was issued by the Nevada State Health Division (NSHD) that recommends limiting human consumption of fish from six northern Nevada waters due to elevated methylmercury levels (NSHD 2007, pp. 1-2). In 2008, the Service published an assessment of trace-metal exposure to aquatic biota from historical mine sites in the western Great Basin (Service 2008, pp. 1-59). The study looked at five different streams across the western Great Basin with various levels of mining impacts (Service 2008, p. 11). The authors found low pH and increased concentrations of certain trace-metals in some streams which pose a significant threat to aquatic biota, increased concentrations of trace-metals in stream sediment, and bioaccumulation of trace-metals in macroinvertebrates and fish (Service 2008, pp. 30-33). In November 2006, a perched aquifer in the headwaters of the North Fork Humboldt River began to drain due to deep core drilling at the Big Springs Mine (HydroGeo 2008, p. 62). Sammy Creek, a tributary to the North Fork Humboldt River, and portions of the North Fork have gone dry in both 2007 and 2008 due to the drained aquifer (HydroGeo 2008, p. 50).

Drought has been an important natural disturbance in the western United States since the early Holocene (Cook *et al.* 2004, p. 1017; Mensing *et al.* 2004, pp. 31-37; Yuan *et al.* 2004, pp. 7-9). Cook *et al.* (2004, p. 1016) report the percentage of the western United States in drought conditions has gradually increased over the last century and that the current drought rivals the drought conditions in the 1930's; however, these more recent droughts (*i.e.*, in the last century) pale in comparison to conditions found 700-1,100 years before present in terms of duration and severity. These historic drought conditions likely negatively impacted Columbia spotted frog populations throughout their range. Due to dispersal abilities, metapopulation dynamics, and unimpaired connected habitat in which they evolved, Columbia spotted frogs were able to persist and repopulate areas when conditions became favorable, despite these severe recurring drought conditions (Lake 2003, pp. 1166-1167; Wilcox *et al.* 2006, p. 859). Since most populations are now isolated, recolonization after extirpation or input of genetic material from other populations cannot occur naturally. With more frequent and severe droughts likely accompanying climate change (see Factor E, Climate Change section below), we conclude that drought is a threat to Columbia spotted frogs throughout their range.

Fire has been one of the dominant factors shaping ecosystems for millennia (Miller and Rose 1999, pp. 555-558). Fire regimes in the Great Basin differ by the three main vegetation types: sagebrush shrublands, desert shrublands, and pinyon-juniper woodlands. Prior to European settlement, fire regimes in sagebrush shrublands of the Great Basin have been characterized as a combination of mixed-severity and stand-replacing fires with return intervals ranging anywhere from 10 to 70 years (Rice *et al.* 2008, p. 154). Desert shrubland vegetation types are characterized by infrequent, stand-replacement fires with fire return intervals between 35 years to several centuries (Rice *et al.* 2008, p. 155). Pinyon-juniper woodlands are characterized as a mixed fire regime; however, fire histories in pinyon-juniper woodlands are difficult to reconstruct (Paysen *et al.* 2000, p. 130). Return intervals in pinyon-juniper woodlands range from 10 to over 300 years depending on site productivity and plant community structure (Rice *et al.* 2008, p. 162). Fire regimes in the Great Basin have become more frequent due to wildfire exclusion, historical grazing practices, and the introduction of invasive nonnative plant species

(Rice *et al.* 2008, p. 141). More frequent fires favor the establishment of nonnative plants (e.g., *Bromus tectorum* Cheatgrass), which results in the loss of sagebrush and other native plant species (Rice *et al.* 2008, p. 154).

Riparian areas are also subject to fires; however, return intervals and fire regimes may be different than the adjacent uplands. The scant information available on fire in riparian areas indicates that return intervals and fire regime type depend on the width of the riparian area and the fuel type adjacent to the riparian area (Dwire and Kauffman 2003, pp. 62-63). Smaller riparian areas are more similar to the adjacent upland areas while larger riparian areas tend to have longer return intervals and lower fire intensity (Dwire and Kauffman 2003, pp. 62-63). Riparian plant species have adapted to disturbances such as fire which, coupled with being in a moist environment, facilitates rapid recovery (Dwire and Kauffman 2003, pp. 67-70).

Changing climate has affected summer temperatures and the timing of spring snowmelt, which have contributed to increasing the length of the wildfire season, wildfire frequency, and the size of wildfires (McKenzie *et al.* 2004, pp. 893-897; Westerling *et al.* 2006, p. 941). Westerling *et al.* (2006, p. 942) conclude that there are robust statistical associations between wildfire and climate in the western United States and that increased fire activity over recent decades reflects responses to climate change (see Factor E, Climate Change section below).

Direct mortality of amphibians due to fire is thought to be rare and of minor importance to most populations (Russell *et al.* 1999, pp. 374-379; Smith 2000, pp. 20, 29-30; Pilliod *et al.* 2003, pp. 165-175; Hossack and Corn 2007, pp. 1406-1409); however, few studies have documented fire effects to aquatic amphibians in the western United States (Bury 2004, pp. 970-973). Most negative effects to aquatic species after wildfire are due to the immediate loss or alteration of habitat and indirect effects such as post-fire hydrologic events (Gresswell 1999, pp. 199-211; Benda *et al.* 2003, pp. 107-117; Miller *et al.* 2003, pp. 121-136; Wondzell and King 2003, pp. 75-84). In addition, fire suppression activities, including construction of fire lines, back burning, application of water from pumps or aerial drops, and use of fire retardants and suppressant foams, could negatively affect amphibians (Little and Calfee 2002, p. 3; Backer *et al.* 2004, pp. 937-944).

In summary, Columbia spotted frog populations have been and continue to be impacted by habitat fragmentation and isolation, interactions with nonnative species, poor habitat condition due to various land use practices, drought, water quality, water management, and fire. Current land uses continue to negatively alter or destroy important habitat throughout the range of the Columbia spotted frog which further fragments populations making them more susceptible to extinction (Wilcox *et al.* 2006, pp. 857-862). Recent advisories pertaining to mercury contamination indicate an increasing risk to populations of Columbia spotted frogs downwind of large mining areas in northeastern Nevada. Based on our evaluation of on-going land use activities described above, we conclude there is sufficient information to develop a proposed listing rule for this species due to the present or threatened destruction, modification, or curtailment of its habitat and range.

B. Overutilization for commercial, recreational, scientific, or educational purposes.

We have no information to support that overutilization is a threat to Great Basin Columbia spotted frogs at this time.

C. Disease or predation.

The impact of non-native invasive species on native species, communities, and ecosystems has been severe (Sakai *et al.* 2001, pp. 305-332). The introduction of non-native salmonid (*Oncorhynchus*, *Salmo*, and *Salvelinus*) and bass (*Micropterus*) species for recreational fishing have negatively affected amphibian species throughout the United States (Bradford 2005, pp. 919-924; Vredenburg and Wake 2007, pp. 5-6), including Columbia spotted frogs (Pilliod and Peterson 2001, pp. 326-331; Tait 2007, pp. 32-33). The negative effects of predation of this kind are difficult to document, particularly in stream systems. However, significant negative effects of predation on frog populations in lentic systems have been documented (Knapp and Matthews 2000, pp. 433-435; Pilliod and Peterson 2001, pp. 326-331; Dunham *et al.* 2004, pp. 19-20; Bradford 2005, pp. 919-924; Knapp 2005, pp. 270-275). In the western United States, Lomnický *et al.* (2007, p. 1086) found that 52 percent of stream lengths surveyed contained nonnative vertebrates. They also found that the most common nonnative vertebrates were brook trout (*Salvelinus fontinalis*) (17 percent of all nonnative vertebrates present), brown trout (*Salmo trutta*) (16 percent), and rainbow trout (*Oncorhynchus mykiss*) (14 percent) (Lomnický *et al.* 2007, p. 1086). Using the same dataset, Whittier and Peck (2008, p. 1889) analyzed the surface area occupied by nonnative vertebrates and found that 75 percent of the waters sampled were occupied by nonnatives. This indicates that there is a greater likelihood of finding nonnative vertebrates in larger streams (Whittier and Peck 2008, p. 1889). When surface area is considered, the most common nonnative vertebrates are rainbow trout, carp (*Cyprinus carpio*), brown trout, and smallmouth bass (*Micropterus dolomieu*) (Whittier and Peck 2008, p. 1890). To date, no State fish and game agencies have altered fish stocking rates or locations in order to benefit Columbia spotted frogs directly; however, conservation efforts for native salmonids may indirectly benefit Columbia spotted frogs due to overlapping distributions.

The bullfrog, a non-native ranid species, occurs within the range of the spotted frog in the Great Basin. Bullfrogs are known to compete with and prey on other frog species (Moyle 1973, pp. 19-21; Pearl *et al.* 2004, pp. 16-18; Monello *et al.* 2006, p. 406; Tait 2007, pp. 32-33). They rarely co-occur with Columbia spotted frogs (one known site in Nevada), but whether this is an artifact of competitive exclusion or predation is unknown at this time. Bullfrogs are important vectors for spreading many types of diseases and parasites to healthy populations of native amphibians (Johnson and Lunde 2005, p. 130).

Although a diversity of microbial species is naturally associated with amphibians, it is generally accepted that they are rarely pathogenic to amphibians except under stressful environmental conditions. Chytridiomycosis (chytrid), caused by the pathogenic fungus *Batrachochytrium dendrobatidis*, is an emerging panzootic fungal disease in the United States and globally (Blaustein *et al.* 2005, pp. 1464-1465; Briggs *et al.* 2005, pp. 3156-3158; Ouellet *et al.* 2005, pp. 1433-1438; Rachowicz *et al.* 2006, pp. 1676-1682; Pounds *et al.* 2006, pp. 161-167; Pearl *et al.* 2007b, pp. 146-148; Vredenburg and Wake 2007, p. 6). Clinical signs of amphibian chytrid and

diagnosis are described by Daszak *et al.* (1999, p. 737) and include abnormal posture, lethargy, and loss of righting reflex. Gross lesions, which are usually not apparent, consist of abnormal epidermal sloughing and ulceration; hemorrhages in the skin, muscle, or eye; hyperemia of digital and ventrum skin, and congestion of viscera. Diagnosis is by identification of characteristic intracellular flask-shaped sporangia and septate thalli within the epidermis. Chytrid can be identified in some species of frogs by examining the oral discs of tadpoles which may be abnormally formed or lacking pigment (Fellers *et al.* 2001, pp. 946-947).

Chytrid was confirmed in the Circle Pond site, Idaho, where long term monitoring since 1998 has indicated a general decline in the population (Engle 2002, p. 15). Sites in both northeast and southeast Oregon have also tested positive for chytrid (Bull 2006, pp. 3-4; Engle 2006, p. 16). Chytrid has also been found in the Wasatch Columbia spotted frog DPS (Wilson *et al.* 2005, pp. 2-3, Semon *et al.* 2005, pp. 11-12). Chytrid has not been found in Columbia spotted frog populations in Nevada; however, chytrid has been found in two bullfrog populations. One of the populations along the Owyhee River in northern Elko County has Columbia spotted frogs (which have not been tested) associated with the infected bullfrogs (D.M. Green, USGS, *in litt.* 2006); the other infected bullfrog population is near Beatty, Nevada, which is approximately 225 km (140 mi) to the south of the Toiyabe Mountains subpopulation (USGS 2005, p. 1). Some evidence suggests that Columbia spotted frogs produce antimicrobial peptides in their skin which may inhibit chytrid infection (Rollins-Smith *et al.* 2002, pp. 473-476; Rollins-Smith *et al.* 2005, pp. 137-142); however, further understanding of how chytrid affects Columbia spotted frogs is needed.

Malformations found in amphibian populations can be caused by several different factors including pesticides, high ultraviolet-B (UV-B) radiation exposure, and parasites and pathogens (Carey *et al.* 2003, pp. 194-197; Ankley *et al.* 2004, pp. 9-13; Johnson and Lunde 2005, pp. 125-138; Sutherland 2005, pp. 109-123). To date, pesticides have not been implicated in Columbia spotted frog declines; UV-B radiation is discussed further below in Factor E. The larvae of the trematode (*Ribeiroia ondatrae*) has been associated with higher than normal levels of malformations in populations of several species of amphibians, including Columbia spotted frogs (Johnson *et al.* 2002, pp. 155-162); however, there is high variability in resistance to infection among amphibian species (Johnson and Hartson 2009, pp. 194-198). Malformed frogs have higher mortality rates than non-malformed individuals (Johnson and Lunde 2005, p. 136). The life cycle of *R. ondatrae* includes three hosts, snails of the genus *Planorbella*, amphibians or fish, and finally a bird or mammal (Johnson and Lunde 2005, p. 126). In a recent study covering five western states, the presence and abundance of *Planorbella* snails was the only variable related to the presence and abundance of *R. ondatrae* (Johnson *et al.* 2002, pp. 160-161). *Planorbella* snails were more associated with wetlands of human origin and higher orthophosphate levels (Johnson *et al.* 2002, pp. 160-161; Johnson and Lunde 2005, pp. 133-135; Johnson *et al.* 2007, pp. 15781-15784). Additionally, two of the four *Planorbella* snail species were recorded at sites beyond their previously known ranges (Johnson *et al.* 2002, p. 161), indicating that this could be an expanding threat to amphibians including Columbia spotted frogs.

In summary, nonnative fish (i.e., salmonids or bass) and amphibian (bullfrog) predators occur

throughout the range of Columbia spotted frogs. These predators can eliminate or reduce populations or restrict movement of individuals thus, increasing fragmentation and not allowing metapopulation dynamics to occur. Nonnative fish and amphibians can also be vectors for parasites or pathogens (*i.e.*, chytrid fungus) which may increase deformities and can increase mortality rates. Based on our evaluation of predation and disease described above, we conclude there is sufficient information to develop a proposed listing rule for this species.

D. The inadequacy of existing regulatory mechanisms.

Spotted frog occurrence sites and potential habitats occur on public, tribal, State, and private lands. This species is included on the Forest Service sensitive species list; as such, its management must be considered during National Forest planning processes. The Forest Service must develop and implement management practices to ensure that species on the sensitive species list do not become threatened or endangered because of Forest Service actions. Management objectives must be met in cooperation with the States when projects on National Forest System lands may have a significant effect on sensitive species population numbers or distributions. Furthermore, for Federal candidate species, management objectives must be implemented in cooperation with the Service.

BLM policies direct management to consider candidate species on public lands under their jurisdiction. Consistent with existing laws, the BLM shall implement management plans that conserve candidate species and their habitats and shall ensure that actions authorized, funded, or carried out by the BLM do not contribute to the need for the species to become listed. Specifically, BLM policy requires the development, cooperation with, and implementation of range-wide or site-specific management plans, conservation strategies, and assessments for candidate species that include specific habitat and population management objectives designed for conservation, as well as management strategies necessary to meet those objectives. The BLM should request technical assistance from the Service, and other qualified sources, on any planned action that may contribute to the need to list a candidate species as threatened or endangered.

Tribal governments within the Great Basin with Columbia spotted frogs do not have regulatory or protective mechanisms in place to protect spotted frogs. The status of local populations of Columbia spotted frogs on Yomba-Shoshone or Duck Valley Shoshone-Paiute Reservation tribal lands is generally unknown.

Columbia spotted frog is classified as a protected amphibian by the State of Nevada under Nevada Administrative Code (NAC) 503.075(3)(a). Per NAC 503.090(1) there is no open season on those species of...amphibian classified as protected. Per NAC 503.093 a person shall not hunt or take any wildlife which is classified as protected, or possess any part thereof, without first obtaining the appropriate license, permit or written authorization from the NDOW. NAC 503.094 authorizes issuance of permits for the take and possession of any species of wildlife for strictly scientific or educational purposes. All Idaho reptiles and amphibians (except bullfrog) are classified as protected non-game species. Protected non-game species status makes it illegal to collect, harm, or otherwise remove from its natural habitat. This designation is held at the

State level to help protect populations. Even though amphibians and reptiles are difficult to maintain in captivity, the rule does allow up to four native amphibians and reptiles of a given species to be captured and held in captivity by holders of a valid Idaho hunting license. Columbia spotted frogs are not on the non-game protected wildlife list for the State of Oregon (635-044-0130). As an indication of its status in the State of Oregon, NatureServe (2009) classifies it as imperiled and vulnerable to extirpation and extinction in the State. All three States include Columbia spotted frogs in their State Wildlife Action Plans as a species of conservation concern (IDFG 2005, p. 71; NDOW 2006, pp. 328-329; ODFW 2006, p. 337).

Protection of wetland habitat from loss of water to irrigation or spring development is difficult because most water in the Great Basin has been allocated to water right applicants based on historical use and spring development has already occurred within much of the known habitat of Columbia spotted frogs. Federal lands may have water rights that are approved for wildlife use, but these rights are often superseded by historical rights upstream or downstream that do not provide for minimum flows. Also, most public lands are managed for multiple use and are subject to livestock grazing, silviculture activities, and recreation uses that may be incompatible with spotted frog conservation without adequate mitigation measures.

The threatened Lahontan cutthroat trout (*Oncorhynchus clarkii henshawi*) (LCT) historically occurred throughout the Nevada portion of Columbia spotted frog's range and their distribution still overlaps in some watersheds. Two Recovery Units for the threatened bull trout (*Salvelinus confluentus*) in northeastern Nevada and southwestern Idaho (Jarvis River Recovery Unit) and eastern Oregon (Malheur River Basin Recovery Unit) overlap Columbia spotted frog habitat. Some recovery efforts and regulatory protection measures for these threatened salmonid species should benefit Columbia spotted frogs in some riverine environments where their habitats overlap.

Lands administered by the Forest Service and BLM are interspersed and surrounded by private parcels on which intensive grazing management, irrigation (diversions), agriculture, and mining activities likely typify the land-use practices. There are generally fewer regulatory mechanisms to address activities on private lands. Grazing of interspersed and surrounding private lands could exacerbate the adverse effects of actions on public lands to Columbia spotted frogs, as described previously. Irrigation, agriculture, and mining practices could dewater streams, create migration barriers, or negatively affect water quality. Ongoing or reasonably foreseeable future activities on private lands within the range of Columbia spotted frogs will continue to affect Columbia spotted frogs and their habitat but the extent of that impact is unknown at this time.

In summary, regulatory mechanisms exist for the Columbia spotted frog; however, consistency in applying these mechanisms is unclear. Although all three States include Columbia spotted frog in their State Wildlife Action Plans as a species of conservation concern, Idaho and Oregon still allow some level of take. Nevada does not allow take of the species without a permit; however, enforcement is lacking and harvest levels are unknown. Federal agency policy requires that management activities do not lead to a trend to list candidate species as threatened or endangered. While policies exist to protect Columbia spotted frogs and their habitat on public lands, there is no mechanism to show the effectiveness of these policies. Other federally listed



species occur within the range of Columbia spotted frogs; however, the extent of this overlap and its effectiveness in protecting Columbia spotted frogs and their habitat is unknown. Private lands could be very important to the conservation of Columbia spotted frogs due to their frequent locations on or near waterways, but protective measures for the species in these areas are generally lacking. Based on our evaluation of the inadequacy of existing regulatory mechanisms described above, we conclude there is sufficient information to develop a proposed listing rule for this species.

E. Other natural or manmade factors affecting its continued existence.

Warming trends seen over the past 50 years in the United States are predicted to continue to increase (Field *et al.* 2007, pp. 626-627); however, the magnitude varies spatially across the continent, is most pronounced during spring and winter months, and has affected daily minimum temperatures more than daily maximum temperatures (Field *et al.* 2007, p. 620). Other effects of climate change include, but are not limited to, changes in types of precipitation (Knowles *et al.* 2006, p. 4557), earlier spring run-off (Stewart *et al.* 2005, p. 1152), longer and more intense fire seasons (Brown *et al.* 2004, pp. 375-385; Westerling *et al.* 2006, pp. 941-942; Bachelet *et al.* 2007, pp. 16-17), and more frequent extreme weather events (Diffenbaugh *et al.* 2005, pp. 15775-15777; Rosenzweig *et al.* 2007, p. 109). These changes in climate and subsequent effects can be attributed to the combined effects of greenhouse gases, sulphate aerosols, and natural external forcing (Karoly *et al.* 2003, p. 1203; Barnett *et al.* 2008, p. 1082).

The Intergovernmental Panel on Climate Change (IPCC) states that of all ecosystems, freshwater ecosystems will have the highest proportion of species threatened with extinction due to climate change (Kundzewicz *et al.* 2007, p. 192). Species with narrow temperature tolerances and cold-water species (e.g., amphibians) will likely experience the greatest effects from climate change, and it is anticipated that populations located at the margins of the species' hydrologic and geographic distributions will be affected first (Bates *et al.* 2008, p. 104). Even in relatively pristine areas (e.g., Yellowstone National Park), biologists are documenting amphibian declines and are linking these declines to long-term, large-scale climatic trends (McMenamin *et al.* 2008, pp. 16988-16990).

Past climate scenarios have shaped Great Basin ecosystems (Tausch *et al.* 2004, pp. 24-40). Great Basin ecosystems and their associated riparian areas are expected to be highly sensitive to any future changes in climate (Sala *et al.* 2000, pp. 1772-1773; Fleishman *et al.* 2004, pp. 248-251; Field *et al.* 2007, pp. 627-630). Ecological consequences of climate change to amphibians may include changes in population dynamics, timing of reproduction, changing geographic range, and more broad community and ecosystem level changes (Hansen *et al.* 2001, pp. 766-773; McCarty 2001, pp. 321-325; Inkley *et al.* 2004, p. 9; Corn 2005, pp. 61-62; Parmesan 2006, pp. 637-669; Rahel and Olden 2008, pp. 522-531). Amphibians are sensitive to changes in precipitation and temperature which may increase the risk of extinction for this group of organisms (Boone *et al.* 2003, pp. 131-136; Corn 2005, pp. 59-64; Noss *et al.* 2006, p. 236; Pounds *et al.* 2007, pp. 19-20; Vredenburg and Wake 2007, pp. 6-7).

Increases in UV-B radiation from depletion of stratospheric ozone have been suggested as a

possible threat to amphibian populations (Blaustein *et al.* 1997, pp. 13735-13736; Adams *et al.* 2005, pp. 493-498; Blaustein and Belden 2005, pp. 87-88; Bancroft *et al.* 2008, pp. 990-993). UV-B mainly decreases egg survivorship and increases deformities in developing metamorphs (Blaustein *et al.* 1997, pp. 13735-13736). Columbia spotted frogs are a species that could be susceptible to increases in UV-B radiation because they are a basking species and lay their eggs in shallow water. However, Blaustein *et al.* (1999, pp. 1102-1104) found that Columbia spotted frogs in the embryonic stage were resistant to UV-B because of high levels of photolyase. Additionally, Adams *et al.* (2005, p. 497) found ambiguous results on the effects of UV-B on Columbia spotted frogs and suggested that the relationship be investigated further.

Many of the threats discussed above do not act alone. Multiple stressors can alter the effects of other stressors or act synergistically to affect individuals and populations (IPCC 2002, p. 22; Boone *et al.* 2003, pp. 138-143; Westerman *et al.* 2003, pp. 90-91; Opdam and Wascher 2004, pp. 285-297; Vredenburg and Wake 2007, p. 7). For example, Kiesecker and Blaustein (1995, pp. 11050-11051) describe how UV-B acts with a pathogen to increase embryonic mortality above levels shown with either factor alone. Interactions between current land uses and changing climate conditions are expected to cause shifts in populations, communities, and ecosystems (Hansen *et al.* 2001, p. 767) which may make certain species more vulnerable to extinction (IPCC 2002, p. 22). Additionally, chemicals may exist in the environment at sub-lethal levels; however, UV light may increase the toxicity of these chemicals or may increase an individual's susceptibility to infection, disease, or predation (Boone *et al.* 2003, pp. 138-142; Burkhart *et al.* 2003, pp. 116-120; Bancroft *et al.* 2008, pp. 990-993; Rohr *et al.* 2008, pp. 1235-1237).

In summary, climate change has and is expected to continue to affect Great Basin ecosystems; however, predictions are difficult to make (Fleishman *et al.* 2004, pp. 248-251; Botkin *et al.* 2007, pp. 227-234; Field *et al.* 2007, pp. 627-630). Corn (2005, pp. 59-64) describes many consequences of a changing climate to amphibian species. The effects of multiple stressors such as climate change, habitat destruction, and disease needs further research. The current state of small fragmented populations of Columbia spotted frogs in the Great Basin indicates a high probability of populations disappearing (Wilcox *et al.* 2006, pp. 857-862). Protecting or improving Columbia spotted frogs and their habitat so that they can adapt to expected changes in climate may be the most important conservation action (Chambers *et al.* 2004b, pp. 266-268). Based on our evaluation of other natural or manmade factors affecting its continued existence described above, we conclude there is sufficient information to develop a proposed listing rule for this species.

## CONSERVATION MEASURES PLANNED OR IMPLEMENTED

A 10-year CAS was signed in September 2003 (NDOW 2003a, pp. 1-43; 2003b, pp. 1-55) for both the Northeast (Jarbridge-Independence Range and Ruby Mountains) and the Toiyabe Mountains subpopulations in Nevada. Additionally, a Candidate Conservation Agreement with Assurances was completed in 2006 for the Owyhee subpopulation at Sam Noble Springs, Idaho (Service 2006, pp. 1-45). At the end of 2008, 8 percent of the identified tasks listed in the Northeast CAS have been completed and an additional 79 percent of the tasks have been initiated

at some level (NDOW 2009b, p. i). At the end of 2008, 22 percent of the identified tasks listed in the Toiyabe Mountains CAS have been completed and an additional 68 percent of the tasks have been initiated at some level (NDOW 2009a). Implementing the CASs also includes formulating future conservation actions aimed at alleviating threats to the species. For example, adequate habitat was identified as a limiting factor in the Toiyabe Mountains subpopulation. A habitat enhancement project was completed in Fall 2004 which included the construction or augmentation of 22 ponds in Indian Valley Creek (NDOW 2004a, pp. 4-6). Effectiveness monitoring of this habitat enhancement project as well as the effectiveness of the CASs as a conservation tool is ongoing.

To minimize the effects of grazing on Columbia spotted frog habitat, many grazing allotment closures and grazing exclosure projects have been implemented throughout the frog's range including on Cloverdale Creek and Indian Valley Creek (Toiyabe Mountains subpopulation), and Dry Creek and Sam Noble Springs (Owyhee subpopulation), as well as study sites in northeastern Oregon (Bull 2005, pp. 2, 35-36). Effectiveness monitoring of these projects is vital in determining the impacts of grazing on Columbia spotted frogs in these areas and the validity of these management actions in protecting and enhancing Columbia spotted frog habitat. Additional genetic research is being conducted to clarify the boundary between the Northern and Great Basin clades in southeastern Oregon. Active monitoring, research, and habitat improvement projects are occurring or are being planned throughout the range of the Great Basin DPS of Columbia spotted frogs, which are increasing our knowledge of life history characteristics, population fluctuations, genetics, and threats to the species.

## SUMMARY OF THREATS

Small, highly fragmented populations, characteristic of the majority of existing populations of Columbia spotted frogs in the Great Basin, are highly susceptible to extinction processes. Poor management of Columbia spotted frog habitat including water development, improper grazing, mining activities and nonnative species have and continue to contribute to the degradation and fragmentation of habitat. Emerging fungal diseases such as chytridiomycosis and the spread of parasites are contributing factors to Columbia spotted frog population declines throughout portions of its range. Effects of climate change such as drought and stochastic events such as fire often have detrimental effects to small isolated populations and can often exacerbate existing threats. Based on our evaluation of the five listing factors affecting the continued existence of Columbia spotted frogs in the Great Basin described above, we conclude there is sufficient information to develop a proposed listing rule for this species. We find that this DPS is warranted for listing throughout all its range, and, therefore, find that it is unnecessary to analyze whether it is threatened or endangered in a significant portion of its range.

## RECOMMENDED CONSERVATION MEASURES:

- Reduce threats to Columbia spotted frogs and their habitat
- Maintain, enhance, and restore populations of Columbia spotted frogs and their habitat throughout their current and historical range
- Determine genetic status of all existing populations

- Assess the abundance of Columbia spotted frogs, trends, habitat conditions, and existing and potential threats in a consistent manner throughout their range
- Conduct research that directly supports conservation and management of Columbia spotted frogs and their habitats (i.e., UV-B, chytridiomycosis, parasites, global climate change, synergistic threats, habitat enhancement)

## LISTING PRIORITY

THREAT			
Magnitude	Immediacy	Taxonomy	Priority
High	Imminent	Monotypic genus	1
		Species	2
		Subspecies/population	3
	Non-imminent	Monotypic genus	4
		Species	5
		Subspecies/population	6
<b>Moderate</b> to Low	<b>Imminent</b>	Monotypic genus	7
		Species	8
		<b>Subspecies/population</b>	<b>9*</b>
	Non-imminent	Monotypic genus	10
		Species	11
		Subspecies/population	12

Rationale for Listing Priority Number:

### Magnitude:

Threats to the species and its habitat such as habitat modification and fragmentation, nonnative species, inadequate regulatory mechanisms, and climate occur rangewide but at various intensities. Other threats such as disease and mining-related activities impact local populations. Thus, the overall magnitude of threats is moderate (Appendix A).

### Imminence:

Threats to the species' habitat have occurred for over 100 years and continue to threaten the species today, indicating the threats to the species are imminent (Appendix A). Climate change and its associated extreme weather conditions are occurring now and are expected to increase in the future. Risks from mercury are continuing and may be increasing in northeast Nevada. Chytrid fungus is documented in Idaho and Oregon populations; however its impact to those populations is unknown. Above natural levels of malformations due to parasites have been documented in other parts of its range and may be a threat to Columbia spotted frogs in the Great Basin DPS in the future.

Rationale for Change in Listing Priority Number: We are not proposing to change the Listing Priority Number.

Have you promptly reviewed all of the information received regarding the species for the purpose of determining whether emergency listing is needed? Yes

Is Emergency Listing Warranted? No. While most threats to the species are imminent, the threats are affecting the species at varying magnitudes and intensities. The two CASs and the development of candidate conservation agreements with assurances should provide a roadmap towards recovery. Monitoring the effectiveness of these agreements and willingness of the participants to continue implementation will remain a priority. As a candidate species, Columbia spotted frogs are afforded higher protection from Federal land management agencies.

## DESCRIPTION OF MONITORING

Numerous mark-recapture and presence-absence surveys are occurring throughout the range of the Great Basin DPS of Columbia spotted frogs. Monitoring and research is being conducted by Colorado State University, Boise State University, USGS, BLM, Forest Service, Service, IDFG, NDOW, and the Nevada Natural Heritage Program. Annual reports and research papers are obtained by the Nevada Fish and Wildlife Office and summarized for the CNOR. A rangewide Columbia spotted frog meeting (initiated in 2002) is held every 2 years to discuss various research, monitoring, and conservation activities occurring throughout the entire range of the species. The last meeting was held on March 4, 2008, in Boise, Idaho. The next meeting will be in 2010 in Reno, Nevada.

Substantial effort is needed to conserve this species because it is a wide ranging species and occupies diverse habitat. Because of this, there is a need to conduct a mid-level type of monitoring effort as described in the *Amphibian Research and Monitoring Initiative* (Muths *et al.* 2006, pp. 1-77). Mid-level monitoring documents trends in site occupancy that may be the most useful metric for assessing changes in amphibian status (Muths *et al.* 2006, pp. 5-6). Mid-level monitoring was conducted by USGS in southeast Oregon from 2000 to 2003 (Wente *et al.* 2005, pp. 99-106; Adams *et al.* 2006, p. 10). This effort should be reinstituted and expanded to the entire range of Columbia spotted frogs within the Great Basin DPS. In addition to mid-level monitoring, intensive surveys being conducted in southeastern Oregon, southwestern Idaho, northeast and central Nevada must continue. Like most aquatic species, amphibian populations fluctuate yearly due to climate (Corn 2005, p. 60). It is important to track population changes annually and for significant time periods to distinguish between anthropogenic effects to the species and its habitat and natural population fluctuations.

## COORDINATION WITH STATES

Indicate which State(s) (within the range of the species) provided information or comments on the species or latest species assessment:

Nevada, Idaho, and Oregon comprise the extent of all historical and current Columbia spotted frog populations within the Great Basin DPS. The NDOW and IDFG contributed valuable information on the species.

Indicate which State(s) did not provide any information or comments: Oregon

## LITERATURE CITED

- Adams, M.J., B.R. Hossack, R.A. Knapp, P.S. Corn, S.A. Diamond, P.C. Trenham, and D.B. Fagre. 2005. Distribution patterns of lentic breeding amphibians in relation to ultraviolet radiation exposure in western North America. *Ecosystems* 8:488-500.
- Adams, M.J., C.A. Pearl, S. Galvan, B. McCreary, N. Chelgren, T. Graham, and D. Pilliod. 2006. Amphibian research and monitoring initiative. Pacific Northwest Region 2006 Annual Report. U.S. Geological Survey, Forest and Rangeland Ecosystem Science Center, Corvallis, Oregon. November 17, 2006. 21 pp.
- Adams, M.J., C.A. Pearl, S. Galvan, B. McCreary, N. Chelgren, R. Klaver, and D. Pilliod. 2008. Amphibian research and monitoring initiative. Pacific Northwest Region 2008 Annual Report. U.S. Geological Survey, Forest and Rangeland Ecosystem Science Center, Corvallis, Oregon. December 30, 2008. 22 pp.
- Adams, M.J., C.A. Pearl, B. McCreary, S.K. Galvan, S.J. Wessell, W.H. Wente, C.W. Anderson, and A.B. Kuehl. 2009. Short-term effect of cattle exclosures on Columbia spotted frog (*Rana luteiventris*) populations and habitat in northeastern Oregon. *Journal of Herpetology* 43:132-138.
- Amy, W. 2003. 2002 amphibian survey results. Ruby Mountains, Jarbidge, and Mountain City Ranger Districts, Humboldt-Toiyabe National Forest. April 23, 2003. 6 pp.
- Amy, W. 2004. 2003 Northeastern Nevada spotted frog technical team amphibian survey results. Ruby Mountains, Jarbidge, and Mountain City Ranger Districts, Humboldt-Toiyabe National Forest. April 28, 2004. 3 pp.
- Ankley, G.T., S.J. Degitz, S.A. Diamond, and T.J.E. Tietge. 2004. Assessment of environmental stressors potentially responsible for malformations in North American anuran amphibians. *Ecotoxicology and Environmental Safety* 58:7-16.
- Bachelet, D., J.M. Lenihan, and R.P. Neilson. 2007. Wildfires and global climate change: the importance of climate change for future wildfire scenarios in the western United States. Pages 22-41 in K.L. Ebi, G.A. Meehl, D. Bachelet, J.M. Lenihan, and R.P. Neilson, R.R. Twilley, D.F. Boesch, V.J. Coles, D.G. Kimmel, and W.D. Miller (contributors), *Regional impacts of climate change: four case studies in the United States*. Pew Center on Global Climate Change, Arlington, Virginia.
- Backer, D.M., S.E. Jensen, and G.R. McPherson. 2004. Impacts of fire suppression activities on natural communities. *Conservation Biology* 18:937-946.
- Baird, S.F., and C. Girard. 1853. Communication. *Proceedings of the Academy of Natural Sciences Philadelphia* 6:378-379.



- Bancroft, B.A., N.J. Baker, and A.R. Blaustein. 2008. A meta-analysis of the effects of ultraviolet B radiation and its synergistic interactions with pH, contaminants, and disease on amphibian survival. *Conservation Biology* 22:987-996.
- Bates, B.C., Z.W. Kundzewicz, S. Wu, and J.P. Palutikof (editors). 2008. Climate change and water. Technical Paper of the Intergovernmental Panel on Climate Change, IPCC Secretariat, Geneva. 210 pp.
- Behnke, R.J., and M. Zarn. 1976. Biology and management of threatened and endangered western trouts. General Technical Report GTR-RM-28. Fort Collins, Colorado: U.S. Forest Service, Rocky Mountain Forest and Range Experimental Station. 45 pp.
- Belsky, A.J., A. Matzke, and S. Uselman. 1999. Survey of livestock influences on stream and riparian ecosystems in the western United States. *Journal of Soil and Water Conservation* 54:419-431.
- Benda, L., D. Miller, P. Bigelow, and K. Andras. 2003. Effects of post-wildfire erosion on channel environments, Boise River, Idaho. *Forest Ecology and Management* 178:105-119.
- Bishop, C.A., D.C. Cunningham, G.M. Fellers, J.P. Gibbs, B.D. Pauli, and B.B. Rothermel. 2003. Physical habitat and its alteration: a common ground for exposure of amphibians to environmental stressors. Pages 209-241 in G.L. Linder, S.K. Krest, and D.W. Sparling (editors), *Amphibian decline: an integrated analysis of multiple stressor effects*. SETAC Press, Pensacola, Florida.
- Blankinship, M.E., and J.C. Munger. 2005. Columbia spotted frog monitoring in the Owyhee Mountains of southwestern Idaho: year 2004 results. Field Report from a cost share agreement between Boise State University and the Bureau of Land Management, Idaho Department of Fish and Game, and the U.S. Fish and Wildlife Service. Boise, Idaho. 65 pp.
- Blaustein, A.R., and L.K. Belden. 2005. Ultraviolet radiation. Pages 87-88 in M. Lannoo (editor), *Amphibian Declines: The Conservation Status of United States Species*. University of California Press. Berkeley, California.
- Blaustein, A.R., J.M. Kiesecker, D.P. Chivers, and R.G. Anthony. 1997. Ambient UV-B radiation causes deformities in amphibian embryos. *Proceedings of the National Academy of Sciences of the United States of America* 94:13735-13737.
- Blaustein, A.R., J.B. Hays, P.D. Hoffman, D.P. Chivers, J.M. Kiesecker, W.P. Leonard, A. Marco, D.H. Olson, J.K. Reaser, and R.G. Anthony. 1999. DNA repair and resistance to UV-B radiation in western spotted frogs. *Ecological Applications* 9:1100-1105.
- Blaustein, A.R., J.M. Romansic, E.A. Scheessele, B.A. Han, A.P. Pessier, and J.E. Longcore.

2005. Interspecific variation in susceptibility of frog tadpoles to the pathogenic fungus *Batrachochytrium dendrobatidis*. *Conservation Biology* 19:1460-1468.
- Boone, M.D., P.S. Corn, M.A. Donnelly, E.E. Little, and P.H. Niewiarowski. 2003. Physical stressors. Pages 129-151 in G.L. Linder, S.K. Krest, and D.W. Sparling (editors), *Amphibian decline: an integrated analysis of multiple stressor effects*. SETAC Press. Pensacola, Florida.
- Bos, D.H., and J.W. Sites, Jr. 2001. Phylogeography and conservation genetics of the Columbia spotted frog (*Rana luteiventris*; Amphibia, Ranidae). *Molecular Ecology* 10:1499-1513.
- Botkin, D.B., H. Saxe, M.B. Araújo, R. Betts, R.H.W. Bradshaw, T. Cedhagen, P. Chesson, T.P. Dawson, J.R. Etterson, D.P. Faith, S. Ferrier, A. Guisan, A.S. Hansen, D.W. Hilbert, C. Loehle, C. Margules, M. New, M.J. Sobel, and D.R.B. Stockwell. 2007. Forecasting the effects of global warming on biodiversity. *BioScience* 57:227-236.
- Bradford, D.F. 2005. Factors implicated in amphibian population declines in the United States. Pages 915-925 in M. Lannoo (editor), *Amphibian Declines: The Conservation Status of United States Species*. University of California Press. Berkeley, California.
- Bridges, C.M., and R.D. Semlitsch. 2005. Xenobiotics. Pages 89-92 in M. Lannoo (editor), *Amphibian Declines: The Conservation Status of United States Species*. University of California Press. Berkeley, California.
- Briggs, C.J., V.T. Vredenburg, R.A. Knapp, and L.J. Rachowicz. 2005. Investigating the population level effects of chytridiomycosis: an emerging infectious disease of amphibians. *Ecology* 86:3149-3159.
- Brown, T.J., B.L. Hall, and A.L. Westerling. 2004. The impact of twenty-first century climate change on wildland fire danger in the western United States: an application perspective. *Climatic Change* 62:365-388.
- Bull, E.L. 2005. Ecology of the Columbia spotted frog in northeastern Oregon. General Technical Report PNW-GTR-640. Portland, Oregon: U.S. Department of Agriculture, Forest Service, Pacific Northwest Research Station. 46 pp.
- Bull, E.L. 2006. Monitoring populations and the presence of *Batrachochytrium dendrobatidis* in Columbia spotted frogs (*Rana luteiventris*) in northeastern Oregon. 2006 Annual Report. Pacific Northwest Research Station. La Grande, Oregon. 10 pp.
- Bull, E.L., and M.P. Hayes. 2000. Livestock effects on reproduction of the Columbia spotted frog. *Journal of Range Management* 53:291-294.
- Bull, E.L., and M.P. Hayes. 2001. Post-breeding season movements of Columbia spotted frogs (*Rana luteiventris*) in northeastern Oregon. *Western North American Naturalist* 61:119-

- Burkhart, J.G., J.R. Bidwell, D.J. Fort, and S.R. Sheffield. 2003. Chemical stressors. Pages 111-128 in G.L. Linder, S.K. Krest, and D.W. Sparling (editors), *Amphibian decline: an integrated analysis of multiple stressor effects*. SETAC Press. Pensacola, Florida.
- Bury, R.B. 2004. Wildfire, fuel reduction, and herpetofaunas across diverse landscape mosaics in northwestern forests. *Conservation Biology* 18:968-975.
- Carey, C., D.F. Bradford, J.L. Brunner, J.P. Collins, E.W. Davidson, J.E. Londcore, M. Ouellet, A.P. Pessier, and D.M. Schock. 2003. Biotic factors in amphibian population declines. Pages 153-208 in G.L. Linder, S.K. Krest, and D.W. Sparling (editors), *Amphibian decline: an integrated analysis of multiple stressor effects*. SETAC Press. Pensacola, Florida.
- Center for Biological Diversity, J. Goodall, E.O. Wilson, P. Ehrlich, J. Terborgh, N. Eldredge, T. Eisner, R. Hass, B. Kingsolver, C. Bowden, M. Sheen, the Xerces Society, and Biodiversity Conservation Alliance. 2004. Petition to list 225 plants and animals as endangered species under the Endangered Species Act. Submitted to the Secretary of Interior, May 11, 2004.
- Chambers, J.C., R.J. Tausch, J.L. Korfmacher, D. Germanoski, J.R. Miller, and D. Jewett. 2004a. Effects of geomorphic processes and hydrologic regimes on riparian vegetation. Pages 196-231 in J.C. Chambers, and J.R. Miller (editors), *Great Basin Riparian Ecosystems: Ecology, Management, and Restoration*. Island Press, Washington.
- Chambers, J.C., J.R. Miller, D. Germanoski, and D.A. Weixelman. 2004b. Process based approaches for managing and restoring riparian ecosystems. Pages 261-292 in J.C. Chambers, and J.R. Miller (editors), *Great Basin Riparian Ecosystems: Ecology, Management, and Restoration*. Island Press, Washington.
- Cunningham, J.M., A.J.K. Calhoun, and W.E. Glanz. 2007. Pond-breeding amphibian species richness and habitat selection in a beaver-modified landscape. *Journal of Wildlife Management* 71:2517-2526.
- Corn, P.S. 2005. Climate change and amphibians. *Animal Biodiversity and Conservation* 28:59-67.
- Daszak, P., L. Berger, A.A. Cunningham, A.D. Hyatt, D.E. Green, and R. Speare. 1999. Emerging infectious diseases and amphibian population declines. *Emerging Infectious Diseases* 5:735-748.
- Diffenbaugh, N.S., J.S. Pal, R.J. Trapp, and F. Giorgi. 2005. Fine-scale processes regulate the response of extreme events to global climate change. *Proceedings of the National Academy of Sciences* 102:15774-15778.

- Dunham, J.B., D.S. Pilliod, and M.K. Young. 2004. Assessing the consequences of nonnative trout in headwater ecosystems in western North America. *Fisheries* 29:18-26.
- Dwire, K.A., and J.B. Kauffman. 2003. Fire and riparian ecosystems in landscapes of the western USA. *Forest Ecology and Management* 178:61-74.
- Engle, J.C. 2000. Columbia spotted frog Great Basin population (Owyhee Mountains subpopulation) long-term monitoring plan. Final report from a cost share agreement between Boise State University and the Bureau of Land Management. Boise, Idaho. November 2000.
- Engle, J.C. 2001. Columbia spotted frog project: the translocation of 2 male Columbia spotted frogs between breeding sites within an element occurrence in the Owyhee subpopulation of the Great Basin population. Idaho Department of Fish and Game Report. Boise, Idaho. 19 pp.
- Engle, J.C. 2002. Columbia spotted frog Great Basin population (Owyhee subpopulation) long-term monitoring plan. Year 2001 results. Idaho Department of Fish and Game Report. Boise, Idaho. 64 pp.
- Engle, J.C. 2006. Columbia spotted frog (*Rana luteiventris*) 2006 Monitoring Report Dry Creek, Oregon. Prepared for the Vale District Bureau of Land Management. 57 pp.
- Engle, J.C., and J.C. Munger. 2003. Population fragmentation of spotted frogs in the Owyhee Mountains. Bureau of Land Management Technical Bulletin No. 03-8. Boise, Idaho. April 2003. 15 pp.
- Environmental Protection Agency (USEPA). 2006. Mercury and mercury compounds report: 2004 toxics release inventory. U.S. Environmental Protection Agency, Region 9. December 2006. 4 pp. <http://www.epa.gov/enviro>.
- Fellers, G.M., D.E. Green, and J.E. Longcore. 2001. Oral chytridiomycosis in the mountain yellow-legged frog (*Rana muscosa*). *Copeia* 2001:945-953.
- Field, C.B., L.D. Mortsch, M. Brklacich, D.L. Forbes, P. Kovacs, J.A. Patz, S.W. Running, and M.J. Scott. 2007. North America. Pages 617-652 in M.L. Parry, O.F. Canziani, J.P. Palutikof, P.J. van der Linden, and C.E. Hanson (editors), *Climate change 2007: Impacts, adaptation, and vulnerability. Contribution of Working Group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom.
- Fleishman, E., J.B. Dunham, D.D. Murphy, and P.F. Brussard. 2004. Explanation, prediction, and maintenance of native species richness and composition. Pages 232-260 in J.C. Chambers, and J.R. Miller (editors), *Great Basin Riparian Ecosystems: Ecology,*

- Management, and Restoration. Island Press, Washington.
- Funk, W.C., A.E. Greene, P.S. Corn, and F.W. Allendorf. 2005a. High dispersal in a frog suggests that it is vulnerable to habitat fragmentation. *Biology Letters* 1:13-16.
- Funk, W.C., M.S. Blouin, P.S. Corn, B.A. Maxwell, D.S. Pilliod, S. Amish, and F.W. Allendorf. 2005b. Population structure of Columbia spotted frogs (*Rana luteiventris*) is strongly affected by the landscape. *Molecular Ecology* 14:483-496.
- Funk, W.C., C.A. Pearl, H.M. Draheim, M.J. Adams, T.D. Mullins, and S.M. Haig. 2008. Range-wide phylogenetic analysis of the spotted frog complex (*Rana luteiventris* and *Rana pretiosa*) in northwestern North America. *Molecular Phylogenetics and Evolution* 49:198-210.
- Germanoski, D., and J.R. Miller. 2004. Basin sensitivity to channel incision in response to natural and anthropogenic disturbance. Pages 88-123 in J.C. Chambers, and J.R. Miller (editors), *Great Basin Riparian Ecosystems: Ecology, Management, and Restoration*. Island Press, Washington.
- Gibbs, J.P. 2000. Wetland loss and biodiversity conservation. *Conservation Biology* 14:314-317.
- Green, D.M. 2003. The ecology of extinction: population fluctuation and decline in amphibians. *Biological Conservation* 111:331-343.
- Green, D.M., T.F. Sharbel, J. Kearsley, and H. Kaiser. 1996. Postglacial range fluctuation, genetic subdivision and speciation in the western North American spotted frog complex, *Rana pretiosa*. *Evolution* 50:374-390.
- Green, D.M., H. Kaiser, T.F. Sharbel, J. Kearsley, and K.R. McAllister. 1997. Cryptic species of spotted frogs, *Rana pretiosa* complex, in Western North America. *Copeia* 1997:1-8.
- Gresswell, R.E. 1999. Fire and aquatic ecosystems in forested biomes of North America. *Transactions of the American Fisheries Society* 128:193-221.
- Hansen, A.J., R.P. Neilson, V.H. Dale, C.H. Flather, L.R. Iverson, D.J. Currie, S. Shafer, R. Cook, and P.J. Bartlein. 2001. Global change in forests: responses of species, communities, and biomes. *BioScience* 51:765-779.
- Hatch, K., C.R. Tracy, J.K. Reaser, and S. Blomquist. 2002. Status of the Columbia spotted frog (*Rana luteiventris*) on U.S. Forest Service land in the Toiyabe Mountains, NV. University of Nevada, Reno, Reno, Nevada. 64 pp.
- Higgins, D.K., M.K. Collins, and C.D. Knightes. 2007. Assessment of impacts to trust resources from atmospheric mercury deposition on Ruby Lake National Wildlife Refuge,

- Research Proposal. Department of the Interior, U.S. Fish And Wildlife Service, Region 8. March 30, 2007. 33 pp.
- Hossack, B.R., and P.S. Corn. 2007. Responses of pond-breeding amphibians to wildfire: short-term patterns in occupancy and colonization. *Ecological Applications* 17:1403-1410.
- Hovingh, P. 1990. Investigations of aquatic resources in the Great Basin and adjacent regions with respect to amphibians, mollusks and leeches: a preliminary report for the tri-state region of Idaho, Nevada, and Utah. March 1990. 12 pp. + appendices.
- Howard, A., and J.C. Munger. 2003. Effects of livestock grazing on the invertebrate prey base and on the survival and growth of larvae of the Columbia spotted frog, *Rana luteiventris*. Bureau of Land Management Technical Bulletin No. 03-7. Boise, Idaho. April 2003. 28 pp.
- HydroGeo. 2008. Draft surface and groundwater hydrology technical report: Big Springs exploration project. Draft Report submitted to Great Basin Ecology, Elko, Nevada. HydroGeo, Incorporated, Crested Butte, Colorado. July, 2008. 79 pp.
- Idaho Department of Fish and Game (IDFG), Idaho Department of Parks and Recreation, Bureau of Land Management, Regions 1 and 4 of U.S. Forest Service, and U.S. Fish and Wildlife Service. 1995. Habitat Conservation Assessment and Conservation Strategy: Spotted Frog (*Rana pretiosa*). (Draft) Idaho State Conservation Effort. 33 pp.
- Idaho Department of Fish and Game (IDFG). 2005. Idaho Comprehensive Wildlife Conservation Strategy. Idaho Conservation Data Center, Idaho Department of Fish and Game, Boise, Idaho. September 2005.  
<http://fishandgame.idaho.gov/cms/tech/CDC/cwcs.cfm>
- Inkley, D.B., M.G. Anderson, A.R. Blaustein, V.R. Burkett, B. Felzer, B. Griffith, J. Price, and T.L. Root. 2004. Global climate change and wildlife in North America. Wildlife Society Technical Review 04-2. The Wildlife Society, Bethesda, Maryland. 26 pp.
- Intergovernmental Panel on Climate Change (IPCC). 2002. Climate Change and Biodiversity. IPCC Technical Paper V. H. Gitay, A. Suárez, T. Watson, and D.J. Dokken (editors), WMO-UNEP. <http://www.ipcc.ch/pub/tpbiodiv.pdf>
- Jansen, A., and M. Healey. 2003. Frog communities and wetland condition: relationships with grazing by domestic livestock along an Australian floodplain river. *Biological Conservation* 109:207-219.
- Jewett, D.G., M.L. Lord, J.R. Miller, and J.C. Chambers. 2004. Geomorphic and hydrologic controls on surface and subsurface flow regimes in riparian meadow ecosystems. Pages 124-161 in J.C. Chambers, and J.R. Miller (editors), *Great Basin Riparian Ecosystems: Ecology, Management, and Restoration*. Island Press, Washington.

- Johnson, P.T.J., and R.B. Hartson. 2009. All hosts are not created equal: explaining differential patterns of malformations in an amphibian community. *Journal of Animal Ecology* 78:191-201.
- Johnson, P.T.J., and K.B. Lunde. 2005. Parasite infection and limb malformations: a growing problem in amphibian conservation. Pages 124-138 in M. Lannoo (editor), *Amphibian Declines: The Conservation Status of United States Species*. University of California Press. Berkeley, California.
- Johnson, P.T.J., K.B. Lunde, E.M. Thurman, E.G. Ritchie, S.N. Wray, D.R. Sutherland, J.M. Kapfer, T.J. Frest, J. Bowerman, and A.R. Blaustein. 2002. Parasite (*Ribeiroia ondatrae*) infection linked to amphibian malformations in the western United States. *Ecological Monographs* 72:151-168.
- Johnson, P.T.J., J.M. Chase, K.L. Dosch, R.B. Hartson, J.A. Gross, D.J. Larson, D.R. Sutherland, and S.R. Carpenter. 2007. Aquatic eutrophication promotes pathogenic infection in amphibians. *Proceedings of the National Academy of Sciences* 104:15781-15786.
- Kauffman, J.B., and W.C. Krueger. 1984. Livestock impacts on riparian plant communities and stream-side management implications, a review. *Journal of Range Management* 37:430-437.
- Kauffman, J.B., W.C. Krueger, and M. Vavra. 1983a. Impacts of cattle on streambanks in northeastern Oregon. *Journal of Range Management* 36:683-685.
- Kauffman, J.B., W.C. Krueger, and M. Vavra. 1983b. Effects of late season cattle grazing on riparian plant communities. *Journal of Range Management* 36:685-691.
- Kiesecker, J.M., and A.R. Blaustein. 1995. Synergism between UV-B radiation and a pathogen magnifies amphibian embryo mortality in nature. *Proceedings of the National Academy of Sciences of the United States of America* 92:11049-11052.
- Knapp, R.A. 2005. Effects of nonnative fish and habitat characteristics on lentic herpetofauna in Yosemite National Park, USA. *Biological Conservation* 121:265-279.
- Knapp, R.A., and K.R. Matthews. 2000. Non-native fish introductions and the decline of the mountain yellow-legged frog from within protected areas. *Conservation Biology* 14:428-438.
- Knowles, N., M.D. Dettinger, and D.R. Cayan. 2006. Trends in snowfall versus rainfall for the western United States, 1949-2004. *Journal of Climate* 19:4545-4559.
- Kundzewicz, Z.W., L.J. Mata, N.W. Arnell, P. Döll, P. Kabat, B. Jiménez, K.A. Miller, T. Oki, Z. Sen, and I.A. Shiklomanov. 2007. Freshwater resources and their management. Pages 174-210 in M.L. Parry, O.F. Canziani, J.P. Palutikof, P.J. van der Linden, and C.E.



- Hanson (editors), Climate change 2007: Impacts, adaptation, and vulnerability. Contribution of Working Group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, United Kingdom.
- Lefcort, H., R.A. Meguire, L.H. Wilson, and W.F. Ettinger. 1998. Heavy metals alter the survival, growth, metamorphosis, and antipredatory behavior of Columbia spotted frog (*Rana luteiventris*) tadpoles. Archives of Environmental Contamination and Toxicology 35:447-456.
- Lingo, H.A., and J.C. Munger. 2003. Progress report July 1, 2002- June 30, 2003: Columbia spotted frog- Great Basin population. Idaho Department of Fish and Game Report October 2003. Boise, Idaho. 69 pp.
- Little, E.E., and Calfee. 2002. Environmental implications of fire-retardant chemicals: project summary. Report to the Forest Service, June, 2002. U.S. Geological Survey, Columbia Environment Research Center, Columbia, Missouri. 9 pp.
- Lohr, K., and A. Moser. 2008. Columbia spotted frog Great Basin population (Owyhee subpopulation) long-term monitoring plan: Year 2008 Results. Idaho Department of Fish and Game Progress Report. Boise, Idaho. October 2008. 21 pp.
- Lomnický, G.A., T.R. Whittier, R.M. Hughes, and D.V. Peck. 2007. Distribution of nonnative aquatic vertebrates in western U.S. streams and rivers. North American Journal of Fisheries Management 27:1082-1093.
- McCarty, J.P. 2001. Ecological consequences of recent climate change. Conservation Biology 15:320-331.
- McKenzie, D., Z. Gedalof, D.L. Peterson, and P. Mote. 2004. Climate change, wildfire, and conservation. Conservation Biology 18:890-902.
- McMenamin, S.K., E.A. Hadly, and C.K. Wright. 2008. Climatic change and wetland desiccation cause amphibian decline in Yellowstone National Park. Proceedings of the National Academy of Sciences 105:16988-16993.
- Meneks, M. 2005a. Columbia spotted frog (*Rana luteiventris*) Northeast Nevada 2004 Forest Service Summary Report. Ruby Mountains-Jarvis and Mountain City Ranger Districts. U.S. Forest Service. January 5, 2005. 7 pp.
- Meneks, M. 2005b. Columbia spotted frog (*Rana luteiventris*) Northeast Nevada 2005 Forest Service Summary Report. Ruby Mountains-Jarvis and Mountain City Ranger Districts. U.S. Forest Service. November 30, 2005. 9 pp.
- Meneks, M. 2006. Columbia spotted frog (*Rana luteiventris*) Northeast Nevada 2006 Forest

- Service Summary Report. Ruby Mountains-Jarbridge and Mountain City Ranger Districts. U.S. Forest Service. November 22, 2006. 11 pp.
- Meneks, M. 2008. Columbia spotted frog (*Rana luteiventris*) Northeast Nevada 2008 Forest Service Summary Report. Ruby Mountains-Jarbridge and Mountain City Ranger Districts. U.S. Forest Service. November 4, 2008. 20 pp.
- Meyer, M. 2008. Columbia spotted frog 2008 Monitoring Report, Dry Creek, Oregon. Prepared for the Vale District Bureau of Land Management. U.S. Fish and Wildlife Service, LaGrande, Oregon. 135 pp.
- Miller, D., C. Luce, and L. Benda. 2003. Time, space, and episodicity of physical disturbance in streams. *Forest Ecology and Management* 178:121-140.
- Miller, G.C. 2004. Interim Report- Mercury wet deposition in Nevada. University of Nevada, Reno. Prepared for U.S. Environmental Protection Agency, Office of Water, Region IX, San Francisco, California. 4 pp.
- Miller, R.F., and J.A. Rose. 1999. Fire history and western juniper encroachment in sagebrush steppe. *Journal of Range Management* 52:550-559.
- Minshall, G.W., S.E. Jensen, and W.S. Platts. 1989. The ecology of stream and riparian habitats of the Great Basin Region: A community profile. *Biological Report* 85:7-24.
- Monello, R.J., J.J. Dennehy, D.L. Murray, and A.J. Wirsing. 2006. Growth and behavioral responses of tadpoles of two native frogs to an exotic competitor, *Rana catesbeiana*. *Journal of Herpetology* 40:403-407.
- Moser, A. 2007. Columbia spotted frog Great Basin population (Owyhee subpopulation) long-term monitoring plan: Year 2007 results at Sam Noble Springs. Idaho Department of Fish and Game DRAFT Report. Boise, Idaho. August 2007. 20 pp.
- Moser, A., and G. Patton. 2006. Columbia spotted frog Great Basin population (Owyhee subpopulation) long-term monitoring plan: Year 2006 results at Sam Noble Springs. Idaho Department of Fish and Game Report. Boise, Idaho. December 2006. 16 pp.
- Moyle, P.B. 1973. Effects of introduced bullfrogs, *Rana catesbeiana*, on the native frogs of the San Joaquin Valley, California. *Copeia* 1973:18-22.
- Munger, J.C. 2003. Critical habitat elements for Columbia spotted frogs in the Owyhee Mountains. Report on a challenge cost share agreement between Boise State University and the Bureau of Land Management. Boise, Idaho. January 10, 2003. 15 pp.
- Munger, J.C., and H.A. Lingo. 2003. Reintroduction of beaver to aid restoration of the spotted frog population in Stoneman Creek. Report on the FY2001 and FY2002 research funded

- by a challenge cost share agreement between Boise State University and the Bureau of Land Management. Boise, Idaho. February 10, 2003. 7 pp.
- Munger, J.C., and K. Oelrich. 2006. Columbia spotted frog Great Basin population (Owyhee subpopulation) long-term monitoring plan: year 2005 results. Field Report from a cost share agreement between Boise State University and the Bureau of Land Management, Idaho Department of Fish and Game, and the U.S. Fish and Wildlife Service. Boise, Idaho. April 4, 2006. 19 pp.
- Munger, J.C., M. Gerber, M. Carroll, K. Madric, and C. Peterson. 1996. Status and habitat associations of the spotted frog (*Rana pretiosa*) in southwestern Idaho. Bureau of Land Management Technical Bulletin No. 96- 1. Boise, Idaho. 112 pp.
- Munger, J.C., M. Gerber, and J. Lauders. 1998. Distribution of the Columbia spotted frog, *Rana luteiventris* in the Dry Creek drainage and other sites in Malheur County, Oregon. Final Report from a cost share agreement between Boise State University and the Bureau of Land Management. Vale, Oregon. February 1, 1998. 67 pp.
- Muths, E., A.L. Gallant, G. Campbell, H. Evan, W.A. Battaglin, D.E. Green, J.S. Staiger, S.C. Walls, M.S. Gunzburger, and R.F. Kearney. 2006. The amphibian research and monitoring initiative (ARMI): 5-year report: U.S. Geological Survey Scientific Investigations Report 2006-5224. 77 pp.
- NatureServe. 2009. NatureServe Explorer: An online encyclopedia of life [web application]. Version 7.1. NatureServe, Arlington, Virginia.  
Available <http://www.natureserve.org/explorer>. (Accessed: March 27, 2009).
- Nevada Department of Wildlife (NDOW). 2003a. Conservation agreement and strategy for the Columbia spotted frog (*Rana luteiventris*)- Great Basin Population Nevada- Toiyabe subpopulation, September 2003. Reno, Nevada. Interagency document. 43 pp.
- Nevada Department of Wildlife (NDOW). 2003b. Conservation agreement and strategy for the Columbia spotted frog (*Rana luteiventris*)- Great Basin Population Nevada- Northeast subpopulation, September 2003. Reno, Nevada. Interagency document. 51 pp.
- Nevada Department of Wildlife (NDOW). 2004a. Toiyabe spotted frog conservation agreement and strategy implementation: 2004 annual report. Southern Region, Las Vegas, Nevada. 13 pp.
- Nevada Department of Wildlife (NDOW). 2004b. Toiyabe spotted frog Great Basin subpopulation long-term monitoring plan. Southern Region, Las Vegas, Nevada. 25 pp.
- Nevada Department of Wildlife (NDOW). 2006. Nevada Wildlife Action Plan. Reno, Nevada. June 23, 2006. 546 pp. + appendices.

- Nevada Department of Wildlife (NDOW). 2009a. DRAFT Toiyabe spotted frog conservation agreement and strategy implementation: 2008 annual report. Southern Region, Las Vegas, Nevada. 11 pp.
- Nevada Department of Wildlife (NDOW). 2009b. Great Basin population of Columbia spotted frog, Northeastern subpopulations: Conservation agreement and strategy implementation 2008 annual report. Reno, Nevada. March 2009. 55 pp.
- Nevada State Health Division (NSHD). 2007. The Nevada State Health Division issues fish consumption advisories for six bodies of water. March 8, 2007. 2 pp.  
<http://health.nv.gov>
- Noss, R., B. Csuti, and M.J. Groom. 2006. Habitat fragmentation. Pages 213-251 in M.J. Groom, G.K. Meffe, and C.R. Carroll (editors), *Principles of Conservation Biology* (Third Edition). Sinauer Associates, Incorporated. Sunderland, Massachusetts.
- Opdam, P., and D. Wascher. 2004. Climate change meets habitat fragmentation: linking landscape and biogeographical scale levels in research and conservation. *Biological Conservation* 117:285-297.
- Ouellet, M., I. Mikaelian, B.D. Pauli, J. Rodrigue, and D.M. Green. 2005. Historical evidence of widespread chytrid infection in North American amphibian populations. *Conservation Biology* 19:1431-1440.
- Oregon Department of Fish and Wildlife (ODFW). 2006. The Oregon Conservation Strategy. Salem, Oregon. February 2006. 375 pp. + appendices.
- Parmesan, C. 2006. Ecological and evolutionary responses to recent climate change. *Annual Review of Ecology, Evolution, and Systematics* 37:637-669.
- Patla, D.A., and C.R. Peterson. 1996. The effects of habitat modification on a spotted frog population in Yellowstone National Park. Pages 16-17 in E.D. Koch, G. Wood, C.R. Peterson, and P.S. Corn (editors), *A summary of the conference on declining and sensitive amphibians in the Rocky Mountains and Pacific Northwest*. Idaho Herpetological Society and U.S. Fish and Wildlife Service, Snake River Basin Office Report, Boise, Idaho. 96 pp.
- Paysen, T.E., R.J. Ansley, J.K. Brown, G.J. Gottfried, S.M. Haase, M.G. Harrington, M.G. Narog, S.S. Sackett, and R.C. Wilson. 2000. Fire in western shrubland, woodland, and grassland ecosystems. Pages 121-159 in K.J. Brown, and J.K. Smith (editors), *Wildland fire and ecosystems: effects of fire on flora*. General Technical Report RMRS-GTR-42-vol. 2. Ogden, UT: U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station. 257 pp.
- Pearl, C.A., M.J. Adams, R.B. Bury, and B. McCreary. 2004. Asymmetrical effects of

- introduced bullfrogs (*Rana catesbeiana*) on native ranid frogs in Oregon. *Copeia* 2004:11-20.
- Pearl, C.A., M.J. Adams, and W.H. Wentle. 2007a. Characteristics of Columbia spotted frog (*Rana luteiventris*) oviposition sites in northeastern Oregon, USA. *Western North American Naturalist* 67:86-91.
- Pearl, C.A., E.L. Bull, D.E. Green, J. Bowerman, M.J. Adams, A. Hyatt, and W.H. Wentle. 2007b. Occurrence of the amphibian pathogen *Batrachochytrium dendrobatidis* in the Pacific Northwest. *Journal of Herpetology* 41:145-149.
- Pilliod, D.S., and C.R. Peterson. 2001. Local and landscape effects of introduced trout on amphibians in historically fishless watersheds. *Ecosystems* 4:322-333.
- Pilliod, D.S., C.R. Peterson, and P.I. Ritson. 2002. Seasonal migration of Columbia spotted frogs (*Rana luteiventris*) among complementary resources in a high mountain basin. *Canadian Journal of Zoology* 80:1849-1862.
- Pilliod, D.S., R.B. Bury, E.J. Hyde, C.A. Pearl, and C.S. Corn. 2003. Fire and amphibians in North America. *Forest Ecology and Management* 178:163-181.
- Pounds, J.A., M.R. Bustamante, L.A. Coloma, J.A. Consuegra, M.P.L. Fogden, P.N. Foster, E. La Marca, K.L. Masters, A. Merino-Viteri, R. Puschendorf, S.R. Ron, G.A. Sánchez-Azofeifa, C.J. Still, and B.E. Young. 2006. Widespread amphibian extinctions from epidemic disease driven by global warming. *Nature* 439:161-167.
- Pounds, J.A., A.C.O.Q. Carnaval, and P.S. Corn. 2007. Climate change, biodiversity loss, and amphibian declines. Pages 19-20 in C. Gascon, J.P. Collins, R.D. Moore, D.R. Church, J.E. McKay, and J.R. Mendelson III (editors), *Amphibian Conservation Action Plan. Proceedings: IUCN/SSC Amphibian Conservation Summit 2005*. IUCN/SSC Specialist Group. Gland, Switzerland and Cambridge, United Kingdom. 64 pp.
- Pringle, C. 2006. The fragmentation of aquatic ecosystems and the alteration of hydrologic connectivity. Pages 243-246 in M.J. Groom, G.K. Meffe, and C.R. Carroll (editors), *Principles of Conservation Biology (Third Edition)*. Sinauer Associates, Inc. Sunderland, Massachusetts.
- Rachowicz, L.J., R.A. Knapp, J.A.T. Morgan, M.J. Stice, V.T. Vredenburg, J.M. Parker, and C.J. Briggs. 2006. Emerging infectious disease as a proximate cause of amphibian mass mortality. *Ecology* 87:1671-1683.
- Rahel, F.J., and J.D. Olden. 2008. Assessing the effects of climate change on aquatic invasive species. *Conservation Biology* 22:512-533.
- Reaser, J.K. 1996. Conservation of the spotted frog (*Rana pretiosa*) in Nevada: Multi-scale

- population status and trends assessment. Pages 21-22 in E.D. Koch, G. Wood, C.R. Peterson, and P.S. Corn (editors), A Summary of the Conference on Declining and Sensitive Amphibians in the Rocky Mountains and Pacific Northwest. Idaho Herpetological Society and U.S. Fish and Wildlife Service, Snake River Basin Office Report, Boise, Idaho, February, 1997. 96 pp.
- Reaser, J.K. 1997. Amphibian declines: Conservation science and adaptive management. Doctoral Dissertation. Stanford University. 148 pp.
- Reaser, J.K. 2000. Demographic analyses of the Columbia spotted frog (*Rana luteiventris*): case study in spatiotemporal variation. Canadian Journal of Zoology 78:1158-1167.
- Reaser, J.J., and D.S. Pilliod. 2005. *Rana luteiventris*, Columbia spotted frog. Pages 559-562 in M. Lannoo (editor), Amphibian Declines: The Conservation Status of United States Species. University of California Press, Berkeley.
- Remington, C.L. 1968. Suture-zones of hybrid interaction between recently joined biotas. Pages 321-428 in T. Dobzhansky, M.K. Hecht, and W.C. Steere (editors), Evolutionary Biology. Plenum, New York.
- Rice, P.M., G.R. McPherson, and L.J. Rew. 2008. Fire and nonnative invasive plants in the Interior West Bioregion. Pages 141-173 in K. Zouhar, J.K. Smith, S. Sutherland, and M.L. Brooks (editors), Wildland fire in ecosystems: fire and nonnative invasive plants. General Technical Report RMRS-GTR-42-volume 6. Ogden, Utah: U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station. 355 pp.
- Ripley, E.A., R.E. Redmann, and A.A. Crowder. 1996. Environmental effects of mining. St. Lucie Press, Delray Beach, Florida. 356 pp.
- Rohr, J.R., A.M. Schotthoefer, T.R. Raffel, H.J. Carrick, N. Halstead, J.T. Hoverman, C.M. Johnson, L.B. Johnson, C. Lieske, M.D. Piwoni, P.K. Schoff, and V.R. Beasley. 2008. Agrochemicals increase trematode infections in a declining amphibian species. Nature 455:1235-1239.
- Rollins-Smith, L.A., C. Carey, J. Longcore, J.K. Doersam, A. Boutte, J.E. Bruzgal, and J.M. Conlon. 2002. Activity of antimicrobial skin peptides from ranid frogs against *Batrachochytrium dendrobatidis*, the chytrid fungus associated with global amphibian declines. Developmental and Comparative Immunology 26:471-479.
- Rollins-Smith, L.A., L.K. Reinert, C.J. O'Leary, L.E. Houston, and D.C. Woodhams. 2005. Antimicrobial peptide defenses in amphibian skin. Integrative and Comparative Biology 45:137-142.
- Rosenzweig, C., G. Casassa, D.J. Karoly, A. Imeson, C. Liu, A. Menzel, S. Rawlins, T.L. Root, B. Seguin, and P. Tryjanowski. 2007. Assessment of observed changes and responses in

- natural and managed systems. Pages 79-131 in M.L. Parry, O.F. Canziani, J.P. Palutikof, P.J. van der Linden, and C.E. Hanson (editors), *Climate change 2007: Impacts, adaptation, and vulnerability. Contribution of Working Group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom.
- Russell K.R., D.H. Van Lear, and D.C. Guynn, Jr.. 1999. Prescribed fire effects on herpetofauna: review and management implications. *Wildlife Society Bulletin* 27:374-384.
- Sada, D.W., and G.L. Vinyard. 2002. Anthropogenic changes in biogeography of Great Basin aquatic biota. Pages 277-293 in R. Hershler, D.B. Madsen, and D.R. Currey (editors), *Great Basin Aquatic Systems History*. Smithsonian Contributions to the Earth Sciences, Number 33, Washington D.C.
- Sakai, A.K., F.W. Allendorf, J.S. Holt, D.M. Lodge, J. Molofsky, K.A. With, S. Baughman, R.J. Cabin, J.E. Cohen, N.C. Ellstrand, D.E. McCauley, P. O'Neil, I.M. Parker, J.N. Thompson, and S.G. Weller. 2001. The population biology of invasive species. *Annual Review in Ecology and Systematics* 32:305-332.
- Sala, O.E., F.S. Chapin III, J.J. Armesto, E. Berlow, J. Bloomfield, R. Dirzo, E. Huber-Sanwald, L.F. Huenneke, R.B. Jackson, A. Kinzig, R. Leemans, D.M. Lodge, H.A. Mooney, M. Oesterheld, N.L. Poff, M.T. Sykes, B.H. Walker, M. Walker, and D.H. Wall. 2000. Global biodiversity scenarios for the year 2100. *Science* 287:1770-1774.
- Schmutzer, A.C., M.J. Gray, E.C. Burton, and D.L. Miller. 2008. Impacts of cattle on amphibian larvae and the aquatic environment. *Freshwater Biology* 53:2613-2625.
- Schulz, T.T., and W.C. Leininger. 1990. Differences in riparian vegetation structure between grazed areas and exclosures. *Journal of Range Management* 43:295-299.
- Semlitsch, R.D. 2002. Critical elements for biologically based recovery plans of aquatic-breeding amphibians. *Conservation Biology* 16:619-629.
- Semlitsch, R.D. 2008. Differentiating migration and dispersal processes for pond-breeding amphibians. *Journal of Wildlife Management* 72:260-267.
- Semon, H.L., R.M. Bogardus, K.A. Hatch. 2005. The extent and impact of chytrid infection on Utah's spotted frog (*Rana luteiventris*). Report prepared for the Utah Division of Wildlife Resources. Brigham Young University, Provo, Utah. 15 pp.
- Shovlain, A.M., D.H. Olson, G.M. Riegel, and W.J. Ripple. 2005. Oregon spotted frog (*Rana pretiosa*) habitat use and herbage (or biomass) removal of grazing at Jack Creek, Klamath County, Oregon. M.S. Thesis, Oregon State University, Corvallis, Oregon. 29 pp.

- Smith, J.K. (editor). 2000. Wildland fire in ecosystems: effects of fire on fauna. General Technical Report RMRS-GTR-42-1. Ogden, Utah: U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station. 83 pp.
- Smyth, M. 2004. Final Report – 2004 Spotted Frog Survey Project. Bureau of Land Management, Andrews Resource Area, Burns District, Oregon. September 25, 2004. 11 pp.
- Stebbins, R.C. 2003. A field guide to western reptiles and amphibians. Third Edition. Houghton Mifflin Company. Boston, Massachusetts.
- Stevens, C.E., C.A. Paszkowski, and A.L. Foote. 2007. Beaver (*Castor canadensis*) as a surrogate species for conserving anuran amphibians on boreal streams in Alberta, Canada. *Biological Conservation* 134:1-13.
- Stewart, I.T., D.R. Cayan, and D.M. Dettinger. 2005. Changes toward earlier streamflow timing across the western North America. *Journal of Climate* 18:1136-1155.
- Stuart, S.N., M. Hoffmann, J.S. Chanson, N.A. Cox, R.J. Berridge, P. Ramani, and B.E. Young (editors). 2008. Threatened amphibians of the world. Lynx Edicions, Barcelona, Spain; IUCN, Gland, Switzerland; and Conservation International, Arlington, Virginia, USA.
- Sutherland, D. 2005. Parasites of North American frogs. Pages 109-123 in M. Lannoo (editor), *Amphibian Declines: The Conservation Status of United States Species*. University of California Press, Berkeley.
- Tait, C. 2007. Conservation assessment of the Great Basin population of the Columbia spotted frog. USDA Forest Service, Intermountain Region, Ogden, Utah. 79 pp.
- Tattersall, G.J., and G.R. Ultsch. 2008. Physiological ecology of aquatic overwintering in Ranid frogs. *Biological Reviews* 83:119-140.
- Tausch, R.J., C.L. Nowak, and S.A. Mensing. 2004. Climate change and associated vegetation dynamics during the Holocene: the paleoecological record. Pages 24-48 in J.C. Chambers, and J.R. Miller (editors), *Great Basin Riparian Ecosystems: Ecology, Management, and Restoration*. Island Press, Washington.
- Thompson, H.B. 1913. Description of a new subspecies of *Rana pretiosa* from Nevada. *Proceedings of the Biological Society of Washington* 26:53-56.
- Turner, F.B. 1962. An analysis of geographic variation and distribution of *Rana pretiosa*. *American Philosophical Society Yearbook* 1962. Pp. 325-328.
- U.S. Fish and Wildlife Service (Service). 1989. Endangered and threatened wildlife and plants; 90-day finding on petition to list the spotted frog. October 17, 1989. *Federal Register*



54:42529.

- U.S. Fish and Wildlife Service (Service). 1993. Endangered and threatened wildlife and plants; 12-month finding on petition to list the spotted frog. May 7, 1993. Federal Register 58:27260-27263.
- U.S. Fish and Wildlife Service (Service). 1996. Policy regarding the recognition of Distinct Vertebrate Population Segments under the Endangered Species Act. Federal Register 61:4722-4725. February 7, 1996.
- U.S. Fish and Wildlife Service (Service). 2006. Candidate Conservation Agreement with Assurances for Columbia spotted frog at Sam Noble Springs, Owyhee County, Idaho. Boise, Idaho. 45 pp.
- U.S. Fish and Wildlife Service (Service). 2008. Final Report: Preliminary Assessment of Trace-Metal Exposures to Aquatic Biota from Historical Mine Sites in the Western Great Basin. Project ID: 1130-1F41. Reno, Nevada. 43 pp.
- United States Geological Survey (USGS). 2005. Pathology report on larval bullfrogs from Death Valley National Monument, Nye County, Nevada. United States Geological Survey, National Wildlife Health Center. Madison, Wisconsin. 3 pp.
- United States Geological Survey (USGS). 2006. Genetic evaluation of the Great Basin distinct population segment in Columbia spotted frogs (*Rana luteiventris*). Fiscal Year 2006 SSP Proposal. U.S. Geological Survey, Forest and Rangeland Ecosystem Science Center, Corvallis, Oregon. 6 pp.
- Unrine, J.M., C.H. Jagoe, W.A. Hopkins, and H.A. Brant. 2004. Adverse effects of ecologically relevant dietary mercury exposure in southern leopard frogs (*Rana sphenocephala*) larvae. Environmental Toxicology and Chemistry 23:2964-2970.
- Utah Nature Study Society. 1989. A petition to the U.S. Fish and Wildlife Service to place the spotted frog, *Rana pretiosa*, in general, and the Wasatch population in particular, on the List of Endangered and Threatened wildlife and plants. May 1, 1989. 12 pp.
- Vredenburg, V.T., and B.D. Wake. 2007. Global declines of amphibians. Encyclopedia of Biodiversity 2007:1-9.
- Welch, N.E., and J.A. MacMahon. 2005. Identifying habitat variables important to the rare Columbia spotted frog in Utah (U.S.A.): an information-theoretic approach. Conservation Biology 19:473-481.
- Wells, K.D. 2007. The ecology and behavior of amphibians. The University of Chicago Press, Chicago. 1,147 pp.
- Wente, W.H., M.J. Adams, and C.A. Pearl. 2005. Evidence of decline for *Bufo boreas* and

*Rana luteiventris* in and around the northern Great Basin, western USA. *Alytes* 22:95-108.

Westerling, A.L., H.G. Hidalgo, D.R. Cayan, and T.W. Swetnam. 2006. Warming and earlier spring increase western U.S. forest wildfire activity. *Science* 313:940-943.

Westerman, A.G., W. van der Schalie, S.L. Levine, B. Palmer, D. Shank, and R.G. Stahl. 2003. Linking stressors with potential effects on amphibian populations. Pages 73-109 in G.L. Linder, S.K. Krest, and D.W. Sparling (editors), *Amphibian decline: an integrated analysis of multiple stressor effects*. Pensacola, Florida, USA: Society of Environmental Toxicology and Chemistry.

Whittier, T.R., and D.V. Peck. 2008. Surface area estimates of streams and rivers occupied by nonnative fish and amphibians in the western USA. *North American Journal of Fisheries Management* 28:1887-1893.

Wilcox, C., B.J. Cairns, and H.P. Possingham. 2006. The role of habitat disturbance and recovery in metapopulation persistence. *Ecology* 87:855-863.

Wilson, K.W., M.D. Mills, E.M. DuRoss. 2005. Columbia spotted frog (*Rana luteiventris*) chytrid monitoring summary. Utah Division of Wildlife Resources, Central Region. Springville, Utah. 5 pp.

Wondzell, S.M., and J.G. King. 2003. Postfire erosional processes in the Pacific Northwest and Rocky Mountain regions. *Forest Ecology and Management* 178:75-87.

Young, B.E., S.N. Stuart, J.S. Chanson, N.A. Cox, and T.M. Boucher. 2004. *Disappearing Jewels: The status of New World amphibians*. NatureServe, Arlington, Virginia. 54 pp.

#### *In Litt.* References


Green, David. 2006. U.S. Geological Survey, National Wildlife Health Center, Madison, Wisconsin. RE: Prelim Findings on larval bullfrogs from Owyhee River, Elko Co. E-mail to Chris Drake, Nevada Department of Wildlife, Elko, Nevada. Dated August 30, 2006.

#### Personal Communication

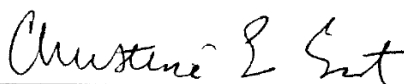
Petersen, Jeff. 2009. Fisheries Biologist, Eastern Region, Nevada Department of Wildlife, Elko, Nevada. Electronic mail received on March 30, 2009. Subject: Columbia spotted frog sampling.

APPROVAL/CONCURRENCE: Lead Regions must obtain written concurrence from all other Regions within the range of the species before recommending changes, including elevations or removals from candidate status and listing priority changes; the Regional Director must approve all such recommendations. The Director must concur on all resubmitted 12-month petition findings, additions or removal of species from candidate status, and listing priority changes.

Approve:

  
Regional Director, Fish and Wildlife Service

6/15/09  
Date

  
**Acting**  
Director, Fish and Wildlife Service

Concur:

Date October 29, 2009

Do not concur: \_\_\_\_\_  
Director, Fish and Wildlife Service

Date

Director's Remarks:

Date of annual review: April 15, 2009  
Conducted by: Chad Mellison

FY 2009, R8 Columbia spotted frog